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(54) Title: MALTOGENIC ALPHA-AMYLASE VARIANTS

(57) Abstract

The inventors have modified the amino acid sequence of a maltogenic alpha-amylase to obtain variants with improved properties, based on the three-dimensional structure of the maltogenic alpha-amylase Novamyl. The variants have altered physicochemical properties, e.g. an altered phyoptimum, improved thermostability, increased specific activity, an altered cleavage pattern or an increased ability to reduce retrogradation of starch or staling of bread.

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MALTOGENIC ALPHA-AMYLASE VARIANTS

FIELD OF THE INVENTION

The present invention relates to variants of maltogenic amylase and to methods of constructing such variants.

5 BACKGROUND OF THE INVENTION

Maltogenic alpha-amylase (glucan 1,4- α -maltohydrolase, E.C. 3.2.1.133) is able to hydrolyze amylose and amylopectin to maltose in the alpha-configuration, and is also able to hydrolyze maltotriose as well as cyclodextrin.

A maltogenic alpha-amylase from *Bacillus* (EP 120 693) is commercially available under the trade name Novamyl® (product of Novo Nordisk A/S, Denmark) and is widely used in the baking industry as an anti-staling agent due to its ability to reduce retrogradation of starch. Novamyl® shares several characteristics with cyclodextrin glucanotransferases (CGTases), including sequence homology (Henrissat B., Bairoch A. 1996) and formation of transglycosylation products (Christophersen, C., et al., 1997, Starch, vol. 50, No. 1, 39-45).

Cyclomaltodextrin glucanotransferase (E.C. 2.4.1.19), also designated cyclodextrin glucanotransferase or cyclodextrin glycosyltransferase, abbreviated herein as CGTase, catalyses the conversion of starch and similar substrates into cyclomaltodextrins via an intramolecular transglycosylation reaction, thereby forming cyclomaltodextrins (or CD) of various sizes.

CGTases are widely distributed and from several different bacterial sources, Clostridium, Corynebacterium, Bacillus. Brevibacterium, Klebsiella. including Thermoanaerobacter and Thermoanaerobacterium have Micrococcus. extensively described in the literature. A CGTase produced by Thermoanaerobacter sp. 25 has been reported in Norman B E, Jørgensen S T; Denpun Kagaku 1992 39 99-106, and WO 89/03421, and the amino acid sequence has been disclosed in WO 96/33267. The sequence of CGTases from Thermoanaerobacterium thermosulfurigenes and from Bacillus circulansis available on the Internet (SCOP or PDF home pages) as pdf file 1CIU, and the sequence of a CGTase from B. circulans is available as pdf file 1CDG.

Tachibana, Y., Journal of Fermentation and Bioengineering, 83 (6), 540-548 (1997) describes the cloning and expression of a CGTase. Variants of CGTases have been described by Kim, Y. H., Biochemistry and Molecular Biology International, 41 (2), 227-234 (1997); Sin K-A, Journal of Biotechnology, 32 (3), 283-288 (1994); D Penninga, Biochemistry, 34 (10), 3368-3376 (1995); and WO 96/33267.

Recently, the tertiary structure of several CGTases have been reported. Hofman et al. [Hofman B E, Bender H, Schultz G E; <u>J. Mol. Biol.</u> 1989 **209** 793-800] and Klein & Schulz [Klein C, Schulz G E; J. Mol. Biol. 1991 217 737-750] report the tertiary structure of a CGTase derived from Bacillus circulans Strain 8, Kubota et al. [Kubota M, 5 Matsuura Y, Sakai S and Katsube Y; Denpun Kagaku 1991 38 141-146] report the tertiary structure of a CGTase derived from Bacillus stearothermophilus TC-91, Lawson et al. [Lawson C L, van Montfort R, Strokopytov B, Rozeboom H J, Kalk K H, de Vries G E, Penninga D, Dijkhuizen L, and Dijkstra B W; J. Mol. Biol. 1994 236 590-600] report the tertiary structure of a CGTase derived from Bacillus circulans Strain 251, 10 Strokopytov et al. [Strokopytov B, Penninga D, Rozeboom H J; Kalk K H, Dijkhuizen L and Dijkstra B W; Biochemistry 1995 34 2234-2240] report the tertiary structure of a CGTase derived from Bacillus circulans Strain 251, which CGTase has been complexed with acarbose, an effective CGTase inhibitor, and Knegtel et al. [Knegtel R M A, Wind R D, Rozeboom H J, Kalk K H, Buitelaar R M, Dijkhuizen L and Dijkstra B 15 W; J. Mol. Biol. 1996 256 611-622] report the tertiary structure of a CGTase derived from Thermoanaerobacterium thermosulfurigenes.

BRIEF DISCLOSURE OF THE INVENTION

The inventors have modified the amino acid sequence of a maltogenic alphaamylase to obtain variants with improved properties, based on the three-dimensional structure of the maltogenic alpha-amylase Novamyl. The variants have altered physicochemical properties., e.g. an altered pH optimum, improved thermostability, increased specific activity, an altered cleavage pattern or an increased ability to reduce retrogradation of starch or staling of bread.

Accordingly, the present invention provides a method of constructing a variant of a parent maltogenic alpha-amylase, wherein the variant has at least one altered property as compared to said parent maltogenic alpha-amylase, which method comprises:

- i) analyzing the structure of the maltogenic alpha-amylase to identify, on the basis of an evaluation of structural considerations, at least one amino acid residue or at least one structural region of the maltogenic alpha-amylase, which is of relevance for altering said property;
 - ii) constructing a variant of the maltogenic alpha-amylase, which as compared to the parent, has been modified in the amino acid residue or structural part identified in i) so as to alter said property; and
 - iii) testing the resulting maltogenic alpha-amylase variant for said property.

The property which may be altered by the above methods of the present invention may be, e.g., stability, pH dependent activity, ability to reduce retrogradation

of starch or staling of bread, specific activity, or substrate specificity. Thus, the variant may have, e.g., increased thermostability or higher activity at a lower pH an altered pH optimum, improved thermostability, increased specific activity or increased ability to reduce retrogradation of starch or staling of bread

In still further aspects the invention relates to variants of a maltogenic alphaamylase, the DNA encoding such variants and methods of preparing the variants. Finally, the invention relates to the use of the variants for various industrial purposes, in particular baking.

DETAILED DISCLOSURE OF THE INVENTION

10 Maltogenic alpha-amylase

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The maltogenic alpha-amylase is an enzyme classified in EC 3.2.1.133. The enzymatic activity does not require a non-reducing end on the substrate and the primary enzymatic activity results in the degradation of amylopectin and amylose to maltose and longer maltodextrins. It is able to hydrolyze amylose and amylopectin to maltose in the alpha-configuration, and is also able to hydrolyze maltotriose as well as cyclodextrin.

A particularly preferred maltogenic alpha-amylase is the amylase cloned from Bacillus as described in EP 120 693 (hereinafter referred to as Novamyl). Novamyl has the amino acid sequence set forth in amino acids 1-686 of SEQ ID NO: 1. Novamyl is encoded in the gene harbored in the Bacillus strain NCIB 11837 which has the nucleic acid sequence set forth in SEQ ID NO:1. The three-dimensional structure of Novamyl is described below.

In general, a preferred maltogenic alpha-amylase should have one or more of the following properties:

- i) a three dimensional structural homology to Novamyl,
- ii) an amino acid sequence having at least 70 % identity to SEQ ID NO: 1, preferably at least 80 % or 90 %, e.g. 95 % or 98 %,
- iii) a DNA sequence which hybridizes to the DNA sequence set forth in SEQ ID NO:1 or to the DNA sequence encoding Novamyl harbored in the *Bacillus* strain NCIB 11837;
- iv) a calcium binding site comprising a coordination equivalent to a backbone carbonyl atom from Asn77, sidechain atom OE2 and OE1 from Glu102, a sidechain atom OD1 from Asp79, a sidechain atom OD1 from Asp76, and a sidechain atom OE1 from Glu101, and one water molecule WAT V21, atom OW0, wherein the positions are as shown in Appendix 1;

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v) a sequence of five amino acids corresponding to Pro-Ala-Gly-Phe-Ser in a position equivalent to residues 191-195 in the amino acid sequence shown in SEQ ID NO: 1; and

The structural homology referred to above in i) is based on other sequence bornologies, hydrophobic cluster analysis or by reverse threading (Huber, T; Torda, AE, PROTEIN SCIENCE Vol. 7, No. 1 pp. 142-149 (1998)) and which by any of these methods is predicted to have the same tertiary structure as Novamyl, wherein the tertiary structure refers to the overall folding or the folding of Domains A, B, and C, more preferably including Domain D, and most preferably including Domain E. Alternatively, a structural alignment between Novamyl and a maltogenic alpha-amylase may be used to identify equivalent positions.

The calcium binding site referred to above in iv) is based on a calcium binding site identified in the three-dimensional structure of Novamyl, and is discussed below in the section "Calcium binding sites."

The "equivalent position" referred to above in v) is based on amino acid or DNA sequence alignment or structural homology using methods known in the art.

Three-dimensional structure of maltogenic alpha-amylase

Novamyl was used to elucidate the three-dimensional structure forming the basis for the present invention.

The structure of Novamyl was solved in accordance with the principle for x-ray crystallographic methods, for example, as given in X-Ray Structure Determination, Stout, G.K. and Jensen, L.H., John Wiley & Sons, Inc. NY, 1989.

The structural coordinates for the solved crystal structure of Novamyl at 2.2 Å resolution using the isomorphous replacement method are given in standard PDB format (Protein Data Bank, Brookhaven National Laboratory, Brookhaven, CT) as set forth in Appendix 1. It is to be understood that Appendix 1 forms part of the present application. In the context of Appendix 1, the following abbreviations are used: CA refers to calcium ion or alpha-carbon atom of the polypeptide backbone, WAT refers to water or to calcium, MAL refers to maltose, HEX refers to a carbohydrate unit of a substrate analogue, and SUL refers to a sulfate ion.

Amino acid residues of the enzyme are identified herein by their respective one- or three-letter amino acid code.

The structure of said maltogenic alpha-amylase is made up of five globular domains, ordered A, B, C, D and E. The domains can be defined as being residues 1-35 132 and 204-403 for Domain A, residues 133-203 for Domain B, residues 404-496 for Domain C, residues 497-579 for Domain D, and residues 580-686 for Domain E,

wherein the numbering refers to the amino acid sequence in SEQ ID NO: 1. Features of Domains A, B, and C of particular interest are described below.

Domain A

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Domain A is the largest domain and contains the active site which comprises a cluster of three amino acid residues, D329, D228 and E256, spatially arranged at the bottom of a cleft in the surface of the enzyme. The structure of Domain A shows an overall fold in common with the α-amylases for which the structure is known, viz. the (beta/alpha) 8 barrel with eight central beta strands (numbered 1-8) and eight flanking a-helices. The β-barrel is defined by McGregor op. cit. The C-terminal end of the beta strand 1 is connected to helix 1 by a loop denoted loop 1 and an identical pattern is found for the other loops, although the loops show some variation in size and some can be quite extensive.

The eight central beta-strands in the (beta/alpha) 8 barrel superimpose reasonably well with the known structures of CGTases. This part of the structure, including the close surroundings of the active site located at the C-terminal end of the beta-strands, shows a high degree of identity with CGTases.

In contrast, the loops connecting the beta-strands and alpha helices display a high degree of variation from the known structures of CGTases. These loops constitute the structural context of the active site, and the majority of the contacts to the substrate is found among residues located in these loops. Distinguishing characteristics such as substrate specificity, substrate binding, pH activity profile, substrate cleavage pattern, and the like, are determined by specific amino acids and the positions they occupy in these loops. In Novamyl Domain A contains two calcium binding sites, one of which is homologous to the calcium binding site in CGTases; the other is unique to Novamyl.

The structure of the calcium binding site is discussed further below in the section "Calcium binding sites."

Domain B

Domain B, also referred to as loop 3 of the (beta/alpha) 8 barrel, in comprises amino acid residues 133-203 of the amino acid sequence shown in SEQ ID NO: 1. The structure is partially homologous to the structure of Domain B in CGTases, the most striking difference being the presence of a five amino acid insert corresponding to positions 191-195 in the amino acid sequence shown in SEQ ID NO: 1 which is not found in the CGTases. This insert is spatially positioned close to the active site residues and in close contact to the substrate.

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Domain C

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Domain C in Novamyl comprises amino acid residues 404-496 of the amino acid sequence shown in SEQ ID NO: 1. Domain C is composed entirely of β-strands which form a single 8-stranded sheet structure that folds back on itself, and thus may be described as a β-sandwich structure. One part of the β-sheet forms the interface to Domain A.

Calcium binding sites

The structure of the maltogenic alpha-amylase exhibits three calcium-binding sites; that is, three calcium ions are found to be present in the structure. In common with most of the known family 13 structures, one calcium ion, WAT 693 in Appendix 1, is located between the A and B domains. This calcium ion is coordinated by a backbone carbonyl atom from Gln184 and His232, sidechain atoms OD2 and OD1 from Asp198, a sidechain atom OD1 from Asn131, and three water molecules WAT V1, WAT V5 and WAT V8.

A second calcium ion is located in the A domain and is common to CGTases, but not found in α -amylases. The calcium ion WAT 694 is coordinated by a backbone carbonyl atom from Gly48 and Asp23, sidechain atom OD2 from Asp50, a sidechain atom OD1 from Asp21, a sidechain atom OD1 from Asp21, and one water molecule WAT V62.

The third calcium ion is located in the A Domain and is unique to Novamyl. The calcium ion is WAT 692 and the coordination comprises a backbone carbonyl atom from Asn77, sidechain atom OE2 and OE1 from Glu102, a sidechain atom OD1 from Asp79, a sidechain atom OD1 from Asp76, and a sidechain atom OE1 from Glu101, and one water molecule WAT V21.

25 Substrate Binding Site

Parts of the loop discussed above in the context of domains A and B are of particular interest for substrate interaction and active site reactivity. In particular, in domain A, residues 37-45 in loop 1, residues 261-266 in loop 5, residues 327-330 in loop 7 and residues 370-376 in loop 8; in domain B, residues 135-145 in loop 3, residues 173-180 and 188-196 in loop 3, wherein residue positions correspond to the amino acids in the amino acid sequence in SEQ ID NO: 1.

Without being limited to any theory, it is presently believed that binding between a substrate and an enzyme is supported by favorable interactions found within a sphere of 4 to 6 Å between the substrate molecule and the enzyme, such as hydrogen bonds and/or strong electrostatic interaction. The following residues of

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Novamyl (SEQ ID NO: 1), are within a distance of 6 Å of the substrate HEX and thus believed to be involved in interactions with said substrate:

44, 89, 90, 92, 93, 127, 129, 132, 135, 177, 178, 188, 191, 194, 196, 226, 228, 229, 230, 231, 232, 256, 258-261, 288, 328, 329, 371, 372, 373, 376, and 690.

The following residues of Novamyl are within a distance of 4 Å of the substrate HEX and thus believed to be involved in interactions with said substrate:

90, 92, 93, 129, 132, 177, 188, 189, 190, 191, 196, 226, 228, 229, 231, 232, 256, 258, 259, 260, 261, 328, 329, 372, 376, and 690.

Homology building of Novamyl®

The structure of the Novamyl® was model built on the structure disclosed in Appendix 1 herein. The structure of other maltogenic alpha-amylases may be built analogously.

A model structure of a maltogenic alpha-amylase can be built using the Homology program or a comparable program, eg., Modeller (both from Molecular Simulations, Inc., San Diego, CA). The principle is to align the sequence of the maltogenic alpha-amylase with the known structure with that of the maltogenic alpha-amylase for which a model structure is to be constructed. The structurally conserved regions can then be built on the basis of consensus sequences. In areas lacking homology, loop structures can be inserted, or sequences can be deleted with subsequent bonding of the necessary residues using, e.g., the program Homology. Subsequent relaxing and optimization of the structure should be done using either Homology or another molecular simulation program, e.g., CHARMm from Molecular Simulations.

Methods for designing novel maltogenic alpha-amylase variants

In a first aspect, the invention relates to a method of constructing a variant of a parent maltogenic alpha-amylase, wherein said variant has at least one altered property as compared to said parent α -amylase, which method comprises:

- i) analyzing the structure of the maltogenic alpha-amylase to identify at least one amino acid or structural region of said α-amylase, which, on the basis of structural or functional considerations, is determined to be of relevance for altering said property of the parent maltogenic alpha-amylase;
 - ii) constructing a variant of the maltogenic alpha-amylase, which as compared to the parent, has been modified in the amino acid residue or structural region identified in i) has been modified so as to alter said property; and
 - iii) testing the resulting variant for said property.

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The structural part which is identified in step i) of the method of the invention may be composed of one amino acid residue. However, normally the structural part comprises more than one amino acid residue, typically constituting one of the above parts of the maltogenic alpha-amylase structure such as one of the A, B, C, D or E domains, an interface between any of these domains, a calcium binding site, a loop structure, the substrate binding site, or the like.

The structural or functional considerations may involve an analysis of the relevant structure or structural part and its contemplated impact on the function of the enzyme. For example, an analysis of the functional differences between maltogenic alpha-amylase and the various CGTases may be used for assigning certain properties of Novamyl to certain parts of the Novamyl structure or to contemplate such relationship. For instance, differences in the pattern or structure of loops surrounding the active site may result in differences in access to the active site of the substrate and thus differences in substrate specificity and/or cleavage pattern.

Furthermore, parts of a maltogenic alpha-amylase involved in substrate binding, and thus, for example, substrate specificity and/or cleavage, calcium ion binding, important, for example, for the calcium dependency of the enzyme, and the like, have been identified (*vide infra*).

The modification of an amino acid residue or structural region is typically accomplished by suitable modifications of a DNA sequence encoding the parent enzyme in question. The modification may be substitution, deletion or insertion of an amino acid residue or a structural part.

The property to be modified may be stability (e.g. thermostability), pH dependent activity, substrate specificity, specific activity or ability to reduce retrogradation of starch or staling of bread. Thus, the altered property may be an altered specific activity at a given pH and/or an altered substrate specificity, such as an altered pattern of substrate cleavage or an altered pattern of substrate inhibition.

In step ii) of the method according to the invention the part of the structure to be identified is preferably one which in the folded enzyme is believed to be in contact with the substrate (cf, the disclosure above in the section entitled "Substrate Binding Site") or involved in substrate specificity and/or cleavage pattern, and/or one which is in contact with one of the calcium ions and/or one, which is contributing to the pH or temperature profile of the enzyme, or is otherwise responsible for the properties of the maltogenic alpha-amylase.

Described in the following are specific types of variants which have been designed by use of the method of the invention.

The variants of the invention may comprise additional modifications in addition to the modifications described herein. The variants preferably have an amino acid

having more than 70 % identity with SEQ ID NO: 1, preferably more than 80 %, particularly more than 90 %, especially more than 95 %, e.g. more than 98 %.

Maltogenic alpha-amylase variants with altered pH dependent activity profile

The pH dependent activity profile can be changed by changing the pKa of residues within 10 Å of the active site residues of the maltogenic alpha-amylase. Changing the pKa of the active site residues is achieved, e.g., by changing the electrostatic interaction or hydrophobic interaction between functional groups of amino acid side chains of a given amino acid residue and its close surroundings. To obtain a higher activity at a higher pH, negatively charged residues are placed near a hydrogen donor acid, whereas positively charged residues placed near a nucleophilic acid will result in higher activity at low pH. Also, a decrease in the pKa can be obtained by reducing the accessibility of water or increasing hydrophobicity of the environment.

Thus, another aspect of the present invention relates to a variant of a parent maltogenic alpha-amylase, in which the variant has an altered pH dependent activity profile as compared to the parent, wherein the variant may be obtained by the following method:

- i) identifying an amino acid residue within 15 Å from an active site residue of a maltogenic alpha-amylase in the three-dimensional structure of said parent maltogenic alpha-amylase, in particular 10 Å from an active site residue, wherein said amino acid residue is contemplated to be involved in electrostatic or hydrophobic interactions with an active site residue;
- ii) substituting, in the structure, said amino acid residue with an amino acid residue which changes the electrostatic and/or hydrophobic surroundings of an active site residue, and evaluating the accommodation of the amino acid residue in the 25 structure.
 - iii) optionally repeating step i) and/or ii) recursively until an amino acid substitution has been identified which is accommodated into the structure,
 - iv) constructing a maltogenic alpha-amylase variant resulting from steps i) and ii), and optionally iii), and testing the pH dependent enzymatic activity of said variant.

In a preferred embodiment, the variant of a maltogenic alpha-amylase having an altered pH dependent activity profile as compared to the parent maltogenic alpha-amylase comprises a modification of an amino acid residue corresponding to one or more of the following residues of the amino acid sequence set forth in SEQ ID NO: 1:

D127, V129, F188, A229, Y258, V281, F284, T288, N327, M330, G370, N371, and D372,

L71, S72, V74, L75, L78, T80, L81, G83, T84, D85, N86, T87, G88, Y89, H90, G91, T94, R95, D96, F97, I174, S175, N176, D178, D179, R180, Y181, E182, A183, Q184, K186, N187, F188, T189, D190, A192, G193, F194, S195, L196.

In more preferred embodiment, the variant comprises a modification 5 corresponding to one or more of the following modifications in the amino acid sequence set forth in SEQ ID NO: 1:

D127N/L, V129S/T/G/V, F188E/K/H, A229S/T/G/V, T288E/K/R, M330L/F/I/D/E/K, F284K/H/D/E/Y. N327D, G370N. V281L/T. N371D/E/G/K, and D372N/V,

L711, S72C, V741, L75N/D/Q/I/V, L78N/I, T80I/L/V/S/N/G, L81I/V/S/T/N/Q/K/H, G83A/S/T/N/Q/E/D/R/H/L, T84S/A/N/D/G, D85A/T/S/N/G, N86Q/E/D/Y/H/K, T87S/I, G88A/S/T, Y89F, H90N/Q/K, G91A/S/T, T94N/D/A/M/V/I, R95K/Q, D96N/V/Q/I, F97Y, 1174N/Q/L, S175T/A/N/D, N176S/T/H/Q/P, D178N/Q/E/K/H, D179Y/N/H, R180W, Y181R/F/C/L. E182D. A183S/C/G, Q184E, K186R. N187Q/E/L/F/H/K/V/L, 15 F188Y/L/I/H/N, T189N/D/A/S/H/Y/G, D190E/Q/H/N/K, A192T/D/E/N/K, G193A/S/T, F194Y, S195N/D/E/R/K/G, L1961.

Similar modifications may be introduced in equivalent positions of other maltogenic alpha-amylases. Variants of particular interest have a combination of one or more of the above with any of the other modifications disclosed herein.

20 Maltogenic alpha-amylase variants with altered stability

A variant with improved stability (typically increased stability) may be obtained by stabilization of calcium binding, substitution with proline, substitution of histidine with another amino acid, introduction of an interdomain disulfide bond, removal of a deamidation site, altering a hydrogen bond contact, filling in an internal structural cavity 25 with one or more amino acids with bulkier side groups, introduction of interdomain interactions, altering charge distribution, helix capping, or introduction of a salt bridge.

Calcium binding

The invention provides a variant of a parent maltogenic alpha-amylase, which has an altered stability due to an altered stabilization of calcium (Ca2+) binding. The 30 enzyme variant may have altered thermostability or pH dependent stability, or it may have maltogenic alpha-amylase activity in the presence of a lower concentration of calcium ion. It is presently believed that amino acid residues located within 10 Å from a calcium ion are involved in or are of importance for the Ca2+ binding capability of the enzyme.

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The amino acid residues found within a distance of 10 Å from the Ca²⁺ binding sites of the maltogenic alpha-amylase with the amino acid sequence set forth in SEQ ID NO: 1 were determined as described in Example 2 and are as follows:

16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28 29, 30, 31, 32, 33,35, 36, 40, 5 46, 47, 48, 49, 50, 51, 52, 53, 54, 56, 73, 74, 75, 76, 77, 78, 79, 80, 81, 87, 88, 89, 91, 93, 94, 95, 96, 99, 100, 101, 102, 103, 104, 105, 109, 129, 130, 131, 132, 133, 134, 145, 150, 167, 168, 169, 170, 171, 172, 174, 177, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 196, 197, 198, 199, 200, 201, 202, 206, 210, 228, 229, 230, 231, 232, 233, 234, 235, 237, 378, and 637.

In order to construct a variant according to this aspect of the invention it is desirable to substitute at least one of the above mentioned amino acid residues, which is determined to be involved in a non-optimal calcium binding, with any other amino acid residue which improves the Ca2+ binding affinity of the variant enzyme. Accordingly, another aspect of the invention relates to a method of constructing a 15 variant of a parent maltogenic alpha-amylase wherein said variant has a stabilised Ca2+ binding as compared to said parent, which method comprises:

- i) identifying an amino acid residue within 10 Å from a Ca2+ binding site of a maltogenic alpha-amylase in a model of the three-dimensional structure of said α amylase which, from structural or functional considerations, is determined to be 20 responsible for a non-optimal calcium ion interaction;
 - ii) constructing a variant in which said amino acid residue is substituted with another amino acid residue which, from structural or functional considerations, is determined to be important for establishing an altered Ca2+ binding affinity; and
 - iii) testing the Ca2+ binding of the resulting maltogenic alpha-amylase variant.

Substituting an amino acid residue responsible for non-optimal calcium ion interaction with another residue may alter a calcium ion binding interaction of the enzyme. For instance, the amino acid residue in question may be selected on the basis of one or more of the following objectives:

- a) to obtain an improved interaction between a calcium ion and an amino acid 30 residue as identified from the structure of the maltogenic alpha-amylase. For instance, if the amino acid residue in question is exposed to a surrounding solvent, it may be advantageous to increase the shielding of said amino acid residue from the solvent so as to stabilize the interaction between said amino acid residue and a calcium ion. This can be achieved by substituting said residue, or an amino acid residue in the vicinity of 35 said residue contributing to the shielding, with an amino acid residue with a bulkier side group or which otherwise results in an improved shielding effect.
 - b) to stabilize a calcium binding site, for instance by stabilizing the structure of the maltogenic alpha-amylase, e.g. by stabilizing the contacts between two or more of

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the five domains or stabilizing one or more of the individual domains as such. This may, e.g., be achieved by providing for a better coordination to amino acid side chains, which may, e.g., be obtained by substituting an N residue with a D residue and/or a Q residue with an E residue, e.g. within 10 Å, and preferably within 3 or 4 Å, of a calcium binding site.

- c) to improve the coordination between the calcium ion and the calcium binding residues, e.g., by improving the interaction between the ion and the coordinating residues or increasing the number of sidechain coordinations by substituting a coordinating water with an amino acid sidechain.
 - d) replace water by a coordinating calcium amino acid residue.

Preferably, the amino acid residue to be modified is located within 8 Å of a Ca²⁺ ion, preferably within 5 Å of a Ca²⁺ ion. The amino acid residues within 8 Å and 5 Å, respectively, may easily be identified by an analogous method used for identifying amino acid residues within 10 Å (cf. Example 2).

In a preferred embodiment, the variant of a maltogenic alpha-amylase having an altered Ca²⁺ binding as compared to the parent maltogenic alpha-amylase comprises a substitution of an amino acid residue corresponding to one or more of the following residues of the amino acid sequence set forth in SEQ ID NO: 1:

D17, A30, S32, R95, H103, N131, Q201, I174, and/or H169,

V74, L75, L78, T80, L81, T87, G88, Y89, H90, G91, T94, R95, D96, F97, Y167, F168, H169, H170, N171, G172, D173, I174, S175, N176, D178, D179, R180, Y181, E182, A183, Q184, K186, N187, F188, T189.

In more preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution corresponding to one or more of the following substitutions in the amino acid sequence set forth in SEQ ID NO: 1:

D17E/Q, A30M/L/A/V/I/E/Q, S32D/E/N/Q, R95M/L/A/V/I/E/Q, H103Y/N/Q/D/E, N131D, Q201E, I174E/Q, and H169N/D/E/Q

V74I, L75N/D/Q/I/V, L78N/I, T80I/L/V/S/N/G, L81I/V/S/T/N/Q/K/H, T87S/I, G88A/S/T, Y89F, H90N/Q/K, G91A/S/T, T94N/D/A/M/V/I, R95K/Q, D96N/V/Q/I, F97Y, Y167F/R/C, F168Y, H169N/Q/K, H170N/Q/K, N171D/E/Q/H/R/K/G, G172A/T/S, D173N/S/T/Y/R/G, I174N/Q/L, S175T/A/N/D, N176S/T/H/Q/P, D178N/Q/E/K/H, D179Y/N/H, R180W, Y181R/F/C/L, E182D, A183S/C/G, Q184E, K186R, N187Q/E/L/F/H/K/V/L, F188Y/L/I/H/N, T189N/D/A/S/H/Y/G.

In another preferred embodiment of the invention with respect to altering the 35 Ca²⁺ binding of a maltogenic alpha-amylase the partial sequence N28-P29-A30-K31-S32-Y33-G34 as set forth in SEQ ID NO: 1 is modified.

Similar substitutions may be introduced in equivalent positions of other maltogenic alpha-amylases. Modifications of particular interest are any combination of one or more of the above with any of the other modifications disclosed herein.

Other substitutions

Variants with improved stability of the enzyme can be achieved by improving existing or introducing new interdomain and intradomain contacts. Such improved stability can be achieved by the modifications listed below.

The maltogenic alpha-amylase having the amino acid sequence shown in SEQ ID NO: 1 may be stabilized by the introduction of one or more interdomain disulfide bonds. Accordingly, another preferred embodiment of the present invention relates to a variant of a parent maltogenic alpha-amylase which has improved stability and at least one more interdomain disulfide bridge as compared to said parent, wherein said variant comprises a modification in a position corresponding to at least one of the following pairs of positions in SEQ ID NO: 1:

G236 + S583, G618 + R272, T252 + V433 and/or A348 + V487.

In a more preferred embodiment, the substitution corresponds to at least one of the following pairs:

G236C + S583C, G618C + R272C, T252C + V433C and/or A348C + V487C.

Another preferred embodiment of the invention relates to a variant of a parent maltogenic alpha-amylase which has an improved stability and an altered interdomain interaction as compared to said parent, wherein said variant comprises a substitution in a position corresponding at least one of the following sets of positions in SEQ ID NO: 1:

- i) F143, F194, L78;
- ii) A341, A348, L398, I415, T439, L464, L465;
- 25 iii) L557;

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- iv) \$240, L268;
- v) Q208, L628;
- vi) F427, Q500, N507, M508, S573; and
- vii) 1510, V620.

In a more preferred embodiment, the substitution corresponds to at least one of the following sets:

- i) F143Y, F194Y, L78Y/F/W/E/Q;
- ii) A341S/D/N, A348V/I/L, L398E/Q/N/D, I415E/Q, T439D/E/Q/N, L464D/E, L465D/E/N/Q/R/K;
 - iii) L557Q/E/N/D;
 - iv) S240D/E/N/Q, L268D/E/N/Q/R/K;
 - v) Q208D/E/Q, L628E/Q/N/D;

vi) F427E/Q/R/K/Y, Q500Y, N507Q/E/D, M508K/R/E/Q, S573D/E/N/Q; and/or vii) I510D/E/N/Q/S, V620D/E/N/Q.

Another preferred embodiment of the invention relates to a variant of a parent maltogenic alpha-amylase which has an improved stability and one or more salt bridges as compared to said parent, wherein said variant comprises a substitution in a position corresponding at least one of the following sets of positions in SEQ ID NO: 1:

N106, N320 and Q624.

In a more preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution corresponding to the following substitutions in the amino acid sequence set forth in SEQ ID NO: 1:

N106R, N320E/D and/or Q624E.

Another embodiment of the invention relates to a variant of a parent maltogenic alpha-amylase which has an improved stability and wherein said variant comprises a substitution in a position corresponding at least one of the following sets of positions in SEQ ID NO: 1:

K40, V74, S141, T142, F188, N234, K249, D261, D261, L268, V279, N342, G397, A403, K425, S442, S479, S493, T494, S495, A496, S497, A498, Q500, K520, A555 and N595.

In a more preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution corresponding to one or more of the following substitutions with proline in the amino acid sequence set forth in SEQ ID NO: 1:

V74P, S141P, N234P, K249P, L268P, V279P, N342P, G397P, A403P, S442P, S479P, S493P, T494P, S495P, A496P, S497P, A498P, Q500P, and/or A555P.

Other preferred substitutions are K40R, T142A, F188I/L, D261G, K425E, 25 K520R, and/or N595I.

Analogously, it may be preferred that one or more histidine residues present in the parent maltogenic alpha-amylase is or are substituted with a non-histidine residues such as Y, V I, L, F, M, E, Q, N, or D. Accordingly, in another preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution of an amino acid residue corresponding to one or more of the following residues of the amino acid sequence set forth in SEQ ID NO: 1:

H103, H220, and H344

In a more preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution corresponding to one or more of the following substitutions in the amino acid sequence set forth in SEQ ID NO: 1:

H103Y/V/I/L/F/Y, H220Y/L/M, and H344E/Q/N/D/Y.

It may be preferred that one or more asparagine or glutamine residues present in the parent maltogenic alpha-amylase is or are substituted with a residue lacking the

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amide on the side chain. Accordingly, in another preferred embodiment, the variant of a Novamyl-like comprises a substitution of an amino acid residue corresponding to one or more of the following residues of the amino acid sequence set forth in SEQ ID NO: 1:

Q13, N26, N77, N86, N99, Q119, N120, N131, N152, N171, N176, N187, S Q201, N203, N234, Q247, N266, N275, N276, N280, N287, Q299, N320, N327, N342, Q365, N371, N375, N401, N436, N454, N468, N474, Q500, N507, N513, Q526, N575, Q581, N621, Q624 and N664.

In more preferred embodiment, the variant of a maltogenic alpha-amylase comprises a substitution corresponding to one or more of the following substitutions in the amino acid sequence set forth in SEQ ID NO: 1:

Q13S/T/A/V/L/I/F/M, N26S/T/A/V/L/I, N77S/T/A/V/L/I, N86S/T/A/V/L/I, N99T/S/V/L, Q119T/S, N120S/T/A/V/L/I, N131S/T/A/V/L/I, N152T/S/V/L, N171Y/D/S/T, Q201S/T/A/V/L/I/F/M, N187S/T/A/V/L/I, N203D/S/T/A/V/L/I, N176S/T/A/V/L/I, Q247S/T/AV/L/I/F/M, N266S/T/A/V/L/I. N275S/T/A/V/L/I, N234S/T/A/V/L/I. 15 N276S/T/AV/L/I, N280S/T/AV/L/I, N287S/T/AV/L/I, Q299L/T/S, N320S/T/AV/L/I, N327S/T/AV/L/I, N342S/T/AV/L/I, Q365S/T/AV/L/I, N371S/T/AV/L/I, N375S/T/AV/L/I, N436S/T/A/V/L/I, N454D/S/T/A/V/L/I, N468D/S/T/A/V/L/I, N401S/T/A/V/L/I, N474D/S/T/AV/L/I, Q500S/T/AV/L/I/F/M, N507S/T/AV/L/I, N513S/T/AV/L/I, Q526 D/S/T/A/V/L/I. N575S/T/A/V/L/I. Q581S/T/AV/L/I/F/M, N621S/T/A/V/L/I 20 Q624S/T/A/V/L/I/F/M and N664D/S/T/A/V/L/I.

Another embodiment of the invention relates to a variant of a parent maltogenic alpha-amylase which has improved stability and improved hydrogen bond contacts as compared to said parent, wherein said variant comprises a modification in a position corresponding to one or more of the following positions in SEQ ID NO: 1:

116, L35, M45, P73, D76, D79, A192, I100, A148, A163+G172, L268, V281, D285, L321, F297, N305, K316, S573, A341, M378, A381, F389, A483, A486, I510, A564, F586, K589, F636, K645, A629, and/or T681.

In a preferred embodiment, the modification corresponds to one or more of the following:

I16T/D/N, L35Q, M45K, P73Q, D76E, D79E/Y, A192S/D/N, I100T/S/D/N/E/Q, A148D/N/E/Q/S/T/R/K, A163Y+G172S/D/N, L268R/K, V281/Q, D285R/K, L321Q, F297N/D/Q/E, N305K/R, K316N/D, S573N/D, A341R/K, M378R/K, A381S/D/N, F389Y, A483S/D/N, A486Q/E, I510R/K, A564S/D/N, F586S/D/N, K589S/D/Q/N, F636Y, K645T, A629N/D/E/Q, and/or T681D/N/E/Q/S.

Similar substitutions may be introduced in equivalent positions of other maltogenic alpha-amylases. Substitutions of particular interest are any combination of one or more of the above with any of the other modifications disclosed herein.

Before actually constructing a maltogenic alpha-amylase variant to achieve any of the above objectives, it may be convenient to evaluate whether or not the contemplated amino acid modification can be accommodated into the maltogenic alpha-amylase structure, e.g. into a model of the three-dimensional structure of the parent maltogenic alpha-amylase.

Maltogenic alpha-amylase variants with altered thermostability and/or altered temperature dependent activity profile

The invention further relates to a variant of a parent maltogenic alpha-amylase, which results from substitution, deletion or insertion of one or more amino acid residues so as to obtain a variant having an altered thermostability or temperature dependent activity profile.

The structure of the maltogenic alpha-amylase contains a number of unique internal cavities which may contain water and a number of crevices. In order to increase the thermostability of the polypeptide it may be desirable to reduce the number or size of cavities and crevices, e.g., by introducing one or more hydrophobic contacts, preferably achieved by introducing amino acids with bulkier side groups in the vicinity or surroundings of the cavity. For instance, the amino acid residues to be modified are those which are involved in the formation of the cavity.

Accordingly, in a further aspect the present invention relates to a method of increasing the thermostability and/or altering the temperature dependent activity profile of a parent maltogenic alpha-amylase, which method comprises:

- i) identifying an internal cavity or a crevice of the parent maltogenic alphaamylase in the three-dimensional structure of said polypeptide;
- ii) substituting, in the structure, one or more amino acid residues in the neighbourhood of the cavity or crevice identified in step i) with another amino acid residue which, from structural or functional considerations, is determined to increase the hydrophobic interaction and to fill out or reduce the size of the cavity or crevice; and
- iii) constructing a variant of the parent maltogenic alpha-amylase resulting from step ii) and testing the thermostability and/or temperature dependent activity of the variant.

The structure identified in Appendix 1 may be used for identifying the cavity or crevice of the parent maltogenic alpha-amylase.

It will be understood that the cavity or crevice is identified by the amino acid residues surrounding said cavity or crevice, and that modification of said amino acid residues are of importance for filling or reducing the size of said cavity or crevice. Preferably, the modification is a substitution with a bulkier amino acid residue, i.e. one with a greater side chain volume. For example, all the amino acids are bulkier than Gly,

whereas Tyr and Trp are bulkier than Phe. The particular amino acid residues referred to below are those which in a crystal structure have been found to flank the cavity or crevice in question.

In a preferred embodiment, the variant of a maltogenic alpha-amylase, in order to fill, either completely or partly, cavities located internally in the structure, comprises a substitution of an amino acid residue corresponding to one or more of the following residues of the amino acid sequence set forth in SEQ ID NO: 1:

L51, L75, L78, G88, G91, T94, V114, I125, V126, T134, G157, L217, S235, G236, V254, V279, V281, L286, V289, I290, V308, L321, I325, D326, L343, F349, S353, I359, I405, L448, Q449, L452, I470, G509, V515, S583, G625, L627, L628 and A670.

L71, S72, V74, L75, L78, T80, L81, G83, T84, D85, N86, T87, G88, Y89, H90, G91, T94, R95, D96, F97, Y167, F168, H169, H170, N171, G172, D173, I174, S175, N176, D178, D179, R180, Y181, E182, A183, Q184, K186, N187, F188, T189, D190, A192, G193, F194, S195, L196.

In a more preferred embodiment, the variant of a maltogenic alpha-amylase comprises one or more substitutions corresponding to the following substitutions in the amino acid sequence set forth in SEQ ID NO: 1:

L217 in combination with L75 (e.g. L217F/Y in combination with L75F/Y), 20 L51W, L75F/Y, L78I, G88AV/T, G91T/S/V/N, T94V/I/L, V114V/I/L, I125L/M/F/Y/W, V126I/L, T134V/I/L/M/F/Y/W, G157AV/I/L, L217V/I/M/F/Y/W, S235I/L/M/F/Y/W, G236AV/I/L/M/F/Y/W, V254I/L/M/F/Y/W, V279M/I/L/F, V281I/L/M/F/Y/W, L286F, V289I/L/R, I290M/L/F, V308I/L/M/F/Y/W, L321I/M/F/Y/W, I325L/M/F/Y/W, D326E/Q, L343M/F/Y/W, F349W/Y, S353V/I/L, I359L/M/F/Y/W, I405M/L/Y/F/W, L448Y, Q449Y, L452M/Y/F/W, I470M/L/F, G509AV/I/L/M/S/T/D/N, V515I/L, S583V/I/L/V, G625AV/I/L/M/F/Y/W, L627M/F/Y, L628M/I/F/Y/W and A670V/I/L/M/F/Y/W,

L71I, S72C, V74I, L75N/D/Q/I/V, L78N/I, T80I/L/V/S/N/G, L81I/V/S/T/N/Q/K/H, G83A/S/T/N/Q/E/D/R/H/L, T84S/A/N/D/G, D85A/T/S/N/G, N86Q/E/D/Y/H/K, T87S/I, G88A/S/T, Y89F, H90N/Q/K, G91A/S/T, T94N/D/A/M/V/I, R95K/Q, D96N/V/Q/I, F97Y, Y167F/R/C, F168Y, H169N/Q/K, H170N/Q/K, N171D/E/Q/H/R/K/G, G172A/T/S, D173N/S/T/Y/R/G, I174N/Q/L, S175T/A/N/D, N176S/T/H/Q/P, D178N/Q/E/K/H, D179Y/N/H, R180W, Y181R/F/C/L, E182D, A183S/C/G, Q184E, K186R, N187Q/E/L/F/H/K/V/L, F188Y/L/I/H/N, T189N/D/A/S/H/Y/G, D190E/Q/H/N/K, A192T/D/E/N/K, G193A/S/T, F194Y, S195N/D/E/R/K/G, L196I.

Similar substitutions may be introduced in equivalent positions of other maltogenic alpha-amylases. Variants of particular interest have a combination of one or more of the above with any of the other modifications disclosed herein.

Maltogenic alpha-amylase variants with an alter dicleavag pattern

One aim of the present invention is to change the degradation characteristics of a maltogenic alpha-amylase. Thus, Novamyl hydrolyzes starch to form predominantly maltose (G2) and a small amount of glucose (G1), but virtually no higher oligosaccharides (G3+). It may be desirable to change this cleavage pattern, e.g. so as to form higher amounts of higher oligosaccharides, such as maltotriose (G3), maltotetraose (G4) and maltopentaose (G5).

A variant of a parent maltogenic alpha-amylase in which the substrate cleavage pattern is altered as compared to said parent may be constructed by a method which to comprises:

- i) identifying the substrate binding area of the parent maltogenic alpha-amylase in a model of the three-dimensional structure, e.g. within a sphere of 4 Å from the substrate binding site as defined in the section above entitled "Substrate Binding Site";
- ii) substituting in the model one or more amino acid residues of the substrate binding area of the cleft identified in i) which is or are believed to be responsible for the cleavage pattern of the parent with another amino acid residue which from structural or functional considerations is believed to result in an altered substrate cleavage pattern, or deleting one or more amino acid residues of the substrate binding area contemplated to introduce favorable interactions to the substrate or adding one or more amino acid residues to the substrate binding area contemplated to introduce favorable interactions to the substrate; and
 - iii) constructing a maltogenic alpha-amylase variant resulting from step ii) and testing the substrate cleavage pattern of the variant.

Accordingly, another aspect of the invention relates to a variant of a parent maltogenic alpha-amylase which has an altered substrate binding site as compared to said parent, which variant comprises a modification in a position corresponding to one or both of the following positions in SEQ ID NO: 1:

V281 and/or A629.

In a preferred embodiment, the variant comprises a modification corresponding to:

V281Q and/or A629N/D/E/Q.

Similar modifications may be introduced in equivalent positions of other maltogenic alpha-amylases. Substitutions of particular interest are any combination of one or both of the above with any of the other modifications disclosed herein.

Maltogenic alpha-amylase variants with improved ability to reduce retrogradation of starch and/or staling of bread

The invention provides maltogenic alpha-amylase variants having improved ability to reduce the retrogradation of starch and/or the staling of bread. Preferred variants comprise a modification at one or more positions corresponding to the following amino acid residues in SEQ ID NO: 1:

A30, K40, N115, T142, F188, T189, P191, A192, G193, F194, S195, D261, N327, K425, K520 and N595.

In a more preferred embodiment, the variant comprises one or more modifications corresponding to the following in SEQ ID NO: 1:

A30D, K40R, N115D, T142A, F188L, T189Y, Δ (191-195), D261G, D261G, N327S, K425E, K520R and N595I.

Determination of residues within 10Å from calcium ions

The coordinates of Appendix 1 are read into the INSIGHT program (BIOSYM Technologies). The spatial coordinates are presented showing the bonds between the atoms. The ions are presented as well as the water atoms. The part of the program package for creating subsets was used to create a 10 Å subset around the calcium ions in the structure by using the command ZONE. All residues identified as having an atom within the designated 10 Å distance from a calcium ion are compiled and listed by using the command LIST MOLECULE. By giving the ions the name "VAT CA" in the coordinate file, a 10 Å sphere around all atoms called "VAT CA" is compiled. The specific residues identified in this manner are given further above in the section entitled "Calcium binding".

25 Determination of cavities

The solved structure of Novamyl with the structural coordinates set forth in Appendix 1 reveals many internal crevices and cavities. When analysing for such cavities the Connolly program is normally used (Lee, B. and Richards, F.M. (1971) J. Mol. Biol. 55:379-400). The program uses a probe with radius to search the external and internal surface of the protein. The smallest crevice observable in this way has the probe radius.

To analyse the solved structure a modified version of the Connolly program included in the program of INSIGHT was used. In the first step, the water molecules and the ions were removed by unmerging these atoms from the solved structure. By using the command MOLECULE SURFACE SOLVENT the solvent accessible surface area was calculated for all atoms and residues using a probe radius of 1.4 Å, and

displayed graphically together with the model of the solved structure. The internal cavities are then seen as dot surfaces with no connections to the external surface.

Suggestions for specific modifications to fill out the cavities are given above in the section entitled "Variants with altered thermostability and/or altered temperature 5 dependent activity profile"). By using the homology built structures or/and comparisons based on sequence alignment, mutations for homologous structures of maltogenic alpha-amylases can be made.

Nomenclature for amino acid modifications

The nomenclature used herein for defining mutations is essentially as described in WO 92/05249. Thus, F188H indicates a substitution of the amino acid F (Phe) in position 188 with the amino acid H (His). V129S/T/G/V indicates a substitution of V129 with S, T, G or V. Δ (191-195) or Δ (191-195) indicates a deletion of amino acids in positions 191-195. 192-A-193 indicates an insertion of A between amino acids 15 192 and 193.

Polypeptide sequence identity

For purposes of the present invention, the degree of identity may be suitably determined according to the method described in Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-45, with the following settings for 20 polypeptide sequence comparison: GAP creation penalty of 3.0 and GAP extension penalty of 0.1. The determination may be done by means of a computer program known such as GAP provided in the GCG program package (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711).

The variants of the invention have an amino acid identity with amino acids 1-686 of SEQ ID NO: 1 of at least 70 %, preferably at least 80 %, e.g. at least 90 %, particularly at least 95 % or at least 98 %.

Hybridization

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Suitable experimental conditions for determining hybridization between a 30 nucleotide probe and a homologous DNA or RNA sequence involves presoaking of the filter containing the DNA fragments or RNA to hybridize in 5x SSC (sodium chloride/sodium citrate, Sambrook, et al., 1989) for 10 min, and prehybridization of the filter in a solution of 5x SSC, 5x Denhardt's solution (Sambrook, et al., 1989), 0.5% SDS and 100 µg/ml of denatured sonicated salmon sperm DNA (Sambrook, et al., 35 1989), followed by hybridization in the same solution containing a random-primed

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(Feinberg, A. P. and Vogelstein, B. (1983) Anal. Biochem. 132:6-13), 32P-dCTP-labeled (specific activity > 1 x 10^9 cpm/µg) probe for 12 hours at ca. 45°C. The filter is then washed twice for 30 minutes in 2x SSC, 0.5% SDS at least 55°C (low stringency), preferably at least 60°C (medium stringency), more preferably at least 65°C 5 (medium/high stringency), more preferably at least 70°C (high stringency), even more preferably at least 75°C (very high stringency).

Molecules which hybridize to the oligonucleotide probe under these conditions are detected by exposure to x-ray film.

Methods of preparing variants of maltogenic alpha-amylases

10 Cloning a DNA sequence encoding a Novamyl-like polypeptide

The DNA sequence encoding a parent maltogenic alpha-amylase may be isolated from any cell or microorganism producing the maltogenic alpha-amylase in question, using various methods well known in the art, for example, from the Bacillus strain NCIB 11837.

First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the maltogenic alpha-amylase to be studied. Then, if the amino acid sequence of the α -amylase is known, homologous, labelled oligonucleotide probes may be synthesised and used to identify maltogenic alpha-amylase-encoding clones from a genomic library prepared 20 from the organism in question. Alternatively, a labelled oligonucleotide probe containing sequences homologous to a known α-amylase gene could be used as a probe to identify maltogenic alpha-amylase-encoding clones, using hybridization and washing conditions of lower stringency.

Another method for identifying maltogenic alpha-amylase-encoding clones 25 involves inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming α-amylase negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for maltogenic alpha-amylase, thereby allowing clones expressing maltogenic alphaamylase activity to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. the phosphoroamidite method described by S.L. Beaucage and M.H. Caruthers (1981) or the method described by Matthes et al. (1984). In the phosphoroamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned 35 in appropriate vectors.

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Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by ligating fragments of synthetic, genomic or cDNA origin, wherein the fragments correspond to various parts of the entire DNA sequence, in accordance with techniques well known in the art. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in US 4,683,202 or R.K. Saiki et al. (1988).

Site-directed Mutagenesis

Once a maltogenic alpha-amylase-encoding DNA sequence has been isolated, and desirable sites for modification identified, modifications may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired modification sites; mutant nucleotides are inserted during oligonucleotide synthesis. In a specific method, a single-stranded gap of DNA, bridging the maltogenic alpha-amylase-encoding sequence, is created in a vector carrying the maltogenic alpha-amylase gene. Then the synthetic nucleotide, bearing the desired modification, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated using T4 ligase. A specific example of this method is described in Morinaga et al. (1984): US 4,760,025 discloses the introduction of oligonucleotides encoding multiple modifications by performing minor alterations of the cassette. However, an even greater variety of modifications can be introduced at any one time by the Morinaga method because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method of introducing modifications into a maltogenic alpha-amylase25 encoding DNA sequences is described in Nelson and Long (1989). It involves a 3-step
generation of a PCR fragment containing the desired modification introduced by using
a chemically synthesized DNA strand as one of the primers in the PCR reactions. From
the PCR-generated fragment, a DNA fragment carrying the modification may be
isolated by cleavage with restriction endonucleases and reinserted into an expression
plasmid.

Random Mutagenesis

Random mutagenesis is suitably performed either as localised or regionspecific random mutagenesis in at least three parts of the gene translating to the amino acid sequence shown in question, or within the whole gene.

The random mutagenesis of a DNA sequence encoding a parent maltogenic alpha-amylase may be conveniently performed by use of any method known in the art.



In relation to the above, a further aspect of the present invention relates to a method for generating a variant of a parent Novamyl-like α -amylase, wherein the variant exhibits increased stability at low pH and at low calcium concentration relative to the parent, the method comprising:

- (a) subjecting a DNA sequence encoding the parent Novamyl-like α -amylase to random mutagenesis,
- (b) expressing the mutated DNA sequence obtained in step (a) in a host cell, and
- (c) screening for host cells expressing a Novamyl-like α -amylase variant which has an altered property relative to the parent Novamyl-like α -amylase.
 - Step (a) of the above method of the invention is preferably performed using doped primers, as described in the working examples herein (*vide infra*).

For instance, the random mutagenesis may be performed by use of a suitable physical or chemical mutagenizing agent, by use of a suitable oligonucleotide, or by subjecting the DNA sequence to PCR generated mutagenesis. Furthermore, the random mutagenesis may be performed by use of any combination of these mutagenizing agents. The mutagenizing agent may, e.g., be one which induces transitions, transversions, inversions, scrambling, deletions, and/or insertions.

Examples of a physical or chemical mutagenizing agent suitable for the present purpose include ultraviolet (UV) irradiation, hydroxylamine, N-methyl-N'-nitro-N-nitrosoguanidine (MNNG), O-methyl hydroxylamine, nitrous acid, ethyl methane sulphonate (EMS), sodium bisulphite, formic acid, and nucleotide analogues. When such agents are used, the mutagenesis is typically performed by incubating the DNA sequence encoding the parent enzyme to be mutagenized in the presence of the mutagenizing agent of choice under suitable conditions for the mutagenesis to take place, and selecting for mutated DNA having the desired properties.

When the mutagenesis is performed by the use of an oligonucleotide, the oligonucleotide may be doped or spiked with the three non-parent nucleotides during the synthesis of the oligonucleotide at the positions which are to be changed. The doping or spiking may be done so that codons for unwanted amino acids are avoided. The doped or spiked oligonucleotide can be incorporated into the DNA encoding the maltogenic alpha-amylase enzyme by any published technique, using e.g. PCR, LCR or any DNA polymerase and ligase as deemed appropriate.

Preferably, the doping is carried out using "constant random doping", in which 35 the percentage of wild-type and modification in each position is predefined. Furthermore, the doping may be directed toward a preference for the introduction of certain nucleotides, and thereby a preference for the introduction of one or more specific amino acid residues. The doping may be made, e.g., so as to allow for the

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introduction of 90% wild type and 10% modifications in each position. An additional consideration in the choice of a doping scheme is based on genetic as well as proteinstructural constraints. The doping scheme may be made by using the DOPE program which, inter alia, ensures that introduction of stop codons is avoided.

When PCR-generated mutagenesis is used, either a chemically treated or nontreated gene encoding a parent maltogenic alpha-amylase enzyme is subjected to PCR under conditions that increase the misincorporation of nucleotides (Deshler 1992; Leung et al., Technique, Vol.1, 1989, pp. 11-15).

A mutator strain of E. coli (Fowler et al., Molec. Gen. Genet., 133, 1974, pp. 10 179-191), S. cereviseae or any other microbial organism may be used for the random mutagenesis of the DNA encoding the maltogenic alpha-amylase by, e.g., transforming a plasmid containing the parent enzyme into the mutator strain, growing the mutator strain with the plasmid and isolating the mutated plasmid from the mutator strain. The mutated plasmid may be subsequently transformed into the expression organism.

The DNA sequence to be mutagenized may be conveniently present in a genomic or cDNA library prepared from an organism expressing the parent maltogenic alpha-amylase. Alternatively, the DNA sequence may be present on a suitable vector such as a plasmid or a bacteriophage, which as such may be incubated with or otherwise exposed to the mutagenising agent. The DNA to be mutagenized may also be 20 present in a host cell either by being integrated in the genome of said cell or by being present on a vector harbored in the cell. Finally, the DNA to be mutagenized may be in isolated form. It will be understood that the DNA sequence to be subjected to random mutagenesis is preferably a cDNA or a genomic DNA sequence.

In some cases it may be convenient to amplify the mutated DNA sequence 25 prior to performing the expression step b) or the screening step c). Such amplification may be performed in accordance with methods known in the art, the presently preferred method being PCR-generated amplification using oligonucleotide primers prepared on the basis of the DNA or amino acid sequence of the parent enzyme.

Subsequent to the incubation with or exposure to the mutagenising agent, the 30 mutated DNA is expressed by culturing a suitable host cell carrying the DNA sequence under conditions allowing expression to take place. The host cell used for this purpose may be one which has been transformed with the mutated DNA sequence, optionally present on a vector, or one which was carried the DNA sequence encoding the parent enzyme during the mutagenesis treatment. Examples of suitable host cells are the 35 following: gram positive bacteria such as Bacillus subtilis, Bacillus licheniformis, Bacillus lentus, Bacillus brevis, Bacillus stearothermophilus, Bacillus alkalophilus, Bacillus amyloliquefaciens, Bacillus coagulans, Bacillus circulans, Bacillus lautus, Bacillus

megaterium, Bacillus thuringiensis, Streptomyces lividans or Streptomyces murinus; and gram negative bacteria such as E. coli.

The mutated DNA sequence may further comprise a DNA sequence encoding functions permitting expression of the mutated DNA sequence.

5 Localized random mutagenesis

The random mutagenesis may be advantageously localised to a part of the parent maltogenic alpha-amylase in question. This may, e.g., be advantageous when certain regions of the enzyme have been identified to be of particular importance for a given property of the enzyme, and when modified are expected to result in a variant having improved properties. Such regions may normally be identified when the tertiary structure of the parent enzyme has been elucidated and related to the function of the enzyme.

The localized, or region-specific, random mutagenesis is conveniently performed by use of PCR generated mutagenesis techniques as described above or any other suitable technique known in the art. Alternatively, the DNA sequence encoding the part of the DNA sequence to be modified may be isolated, e.g., by insertion into a suitable vector, and said part may be subsequently subjected to mutagenesis by use of any of the mutagenesis methods discussed above.

For region-specific random mutagenesis with a view to improving the stability of calcium binding of a parent maltogenic alpha-amylase, codon positions corresponding to the following amino acid residues from the amino acid sequence set forth in SEQ ID NO: 1 may appropriately be targeted:

Residues: Regions:

16-33, 35-36, 40: 16-40

46-54, 56: 46-56

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73-81: 73-81

87-89, 91, 93-96, 99-105, 109: 87-109

129-134, (145, 150): 129-134

167-172, 174, 177, 180-189: 167-189

196-202, 206-210: 196-210

228-235, 237: 228-237

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With a view to achieving improved binding of a substrate, i.e., improved binding of a carbohydrate species, such as amylose or amylopectin, by a maltogenic alphaamylase variant with a modified, e.g. higher, substrate specificity and/or a modified, e.g. higher, specificity with respect to cleavage, i.e. hydrolysis, of the substrate, it appears

that the following codon positions in the following regions of the amino acid sequence shown in SEQ ID NO: 1, may particularly appropriately be targeted for modification by region-specific mutagenesis:

70-97, 127-143, 174-198, 226-233, 255-270, 282-292, 324-331, 370-376.

For region-specific random mutagenesis with a view to altering the substrate specificity and/or the pH dependent activity profile, the following regions of SEQ ID NO: 1 may be targeted: 70-97, 174-198.

The following regions may be targeted with a view to improving the thermostability: 70-109, 167-200.

10 General method for random mutagenesis by use of the DOPE program

The random mutagenesis may be carried out by the following steps:

- 1. Select regions of interest for modification in the parent enzyme
- 2.Decide on mutation sites and nonmutated sites in the selected region
- 3.Decide on which kind of mutations should be carried out, e.g. with respect to the desired stability and/or performance of the variant to be constructed
 - 4. Select structurally reasonable mutations
 - 5. Adjust the residues selected by step 3 with regard to step 4.
 - 6. Analyse by use of a suitable dope algorithm the nucleotide distribution.
- 7.If necessary, adjust the wanted residues to genetic code realism, e.g. taking into account constraints resulting from the genetic code, e.g. in order to avoid introduction of stop codons; the skilled person will be aware that some codon combinations cannot be used in practice and will need to be adapted
 - 8.Make primers

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- 9.Perform random mutagenesis by use of the primers
- 10.Select resulting α -amylase variants by screening for the desired improved properties.

Suitable dope algorithms for use in step 6 are well known in the art. One such algorithm is described by Tomandl, D. et al., 1997, Journal of Computer-Aided Molecular Design 11:29-38. Another algorithm is DOPE (Jensen, LJ, Andersen, KV, 30 Svendsen, A, and Kretzschmar, T (1998) Nucleic Acids Research 26:697-702).

Expression of maltogenic alpha-amylase variants

The construction of the variant of interest is accomplished by cultivating a microorganism comprising a DNA sequence encoding the variant under conditions which are conducive for producing the variant, and optionally subsequently recovering the variant from the resulting culture broth. This is described in detail further below.

According to the invention, a DNA sequence encoding the variant produced by methods described above, or by any alternative methods known in the art, can be expressed, in the form of a protein or polypeptide, using an expression vector which typically includes control sequences encoding a promoter, operator, ribosome binding 5 site, translation initiation signal, and, optionally, a repressor gene or various activator genes.

The recombinant expression vector carrying the DNA sequence encoding an maltogenic alpha-amylase variant of the invention may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector 10 will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e. a vector which exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g. a plasmid, a bacteriophage or an extrachromosomal element, minichromosome or an artificial chromosome. Alternatively, the vector may be one which, when introduced into 15 a host cell, is integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows transcriptional activity in the host cell of choice and may be derived from genes encoding pro-20 teins either homologous or heterologous to the host cell. Examples of suitable promoters for directing the transcription of the DNA sequence encoding a maltogenic alpha-amylase variant of the invention, especially in a bacterial host, are the promoter of the lac operon of E.coli, the Streptomyces coelicolor agarase gene dagA promoters, the promoters of the Bacillus licheniformis α-amylase gene (amyL), the promoters of 25 the Bacillus stearothermophilus maltogenic amylase gene (amyM), the promoters of the Bacillus amyloliquefaciens α-amylase (amyQ), the promoters of the Bacillus subtilis xylA and xylB genes, etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding A. oryzae TAKA amylase, Rhizomucor miehei aspartic proteinase, A. niger neutral α -amylase, A. niger acid stable α -amylase, 30 A. niger glucoamylase, Rhizomucor miehei lipase, A. oryzae alkaline protease, A. oryzae triose phosphate isomerase or A. nidulans acetamidase.

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably connected to the DNA sequence encoding the maltogenic alpha-amylase variant of the 35 invention. Termination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.

The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the *dal* genes from *B. subtilis* or *B. licheniformis*, or one which confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracycline resistance. Furthermore, the vector may comprise *Aspergillus* selection markers such as amdS, argB, niaD and sC, a marker giving rise to hygromycin resistance, or the selection may be accomplished by co-transformation, e.g. as described in WO 91/17243.

While intracellular expression may be advantageous in some respects, e.g. when using certain bacteria as host cells, it is generally preferred that the expression is extracellular. In general, the *Bacillus* α-amylases mentioned herein comprise a preregion permitting secretion of the expressed protease into the culture medium. If desirable, this preregion may be replaced by a different preregion or signal sequence, conveniently accomplished by substitution of the DNA sequences encoding the respective preregions.

The procedures used to ligate the DNA construct of the invention encoding maltogenic alpha-amylase variant, the promoter, terminator and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, are well known to persons skilled in the art (cf., for instance, Sambrook et al. (1989)).

The cell of the invention, either comprising a DNA construct or an expression vector of the invention as defined above, is advantageously used as a host cell in the recombinant production of a maltogenic alpha-amylase variant of the invention. The cell may be transformed with the DNA construct of the invention encoding the variant, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA constructs into the host chromosome may be performed according to conventional methods, e.g. by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

The cell of the invention may be a cell of a higher organism such as a mammal or an insect, but is preferably a microbial cell, e.g. a bacterial or a fungal (including yeast) cell.

Examples of suitable bacteria are gram positive bacteria such as Bacillus subtilis, Bacillus licheniformis, Bacillus lentus, Bacillus brevis, Bacillus stearothermo-

philus, Bacillus alkalophilus, Bacillus amyloliquefaciens, Bacillus coagulans, Bacillus circulans, Bacillus lautus, Bacillus megaterium, Bacillus thuringiensis, or Streptomyces lividans or Streptomyces murinus, or gram negative bacteria such as E.coli. The transformation of the bacteria may, for instance, be effected by protoplast transformation or by using competent cells in a manner known per se.

The yeast organism may favourably be selected from a species of Saccharomyces or Schizosaccharomyces, e.g. Saccharomyces cerevisiae. The filamentous fungus may advantageously belong to a species of Aspergillus, e.g. Aspergillus oryzae or Aspergillus niger. Fungal cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed by regeneration of the cell wall in a manner known per se. A suitable procedure for transformation of Aspergillus host cells is described in EP 238 023.

In a yet further aspect, the present invention relates to a method of producing a maltogenic alpha-amylase variant of the invention, which method comprises cultivating a host cell as described above under conditions conducive to the production of the variant and recovering the variant from the cells and/or culture medium.

The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and obtaining expression of the maltogenic alpha-amylase variant of the invention. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g. as described in catalogues of the American Type Culture Collection).

The maltogenic alpha-amylase variant secreted from the host cells may conveniently be recovered from the culture medium by well-known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulfate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

Testing of maltogenic alpha-amylase variants

Maltogenic alpha-amylase variants produced by any of the methods described above may be tested, either prior to or after purification, for amylolytic activity in a screening assay which measures the ability of the variant to degrade starch. The screening in step 10 in the above-mentioned random mutagenesis method of the invention may be conveniently performed by use of a filter assay based on the following procedure: A microorganism capable of expressing the mutated maltogenic alpha-amylase of interest is incubated on a suitable medium and under suitable conditions for secretion of the enzyme, the medium being covered with two filters comprising a protein-binding filter placed under a second filter exhibiting a low protein binding

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capability. The microorganism is grown on the second, top filter. Subsequent to the incubation, the bottom protein-binding filter comprising enzymes secreted from the microorganism is separated from the second filter comprising the microorganism. The protein-binding filter is then subjected to screening for the desired enzymatic activity, 5 and the corresponding microbial colonies present on the second filter are identified. The first filter used for binding the enzymatic activity may be any protein-binding filter, e.g., nylon or nitrocellulose. The second filter carrying the colonies of the expression organism may be any filter that has no or low affinity for binding proteins, e.g., cellulose acetate or Durapore™.

Screening consists of treating the first filter to which the secreted protein is bound with a substrate that allows detection of the α -amylase activity. The enzymatic activity may be detected by a dye, fluorescence, precipitation, pH indicator, IRabsorbance or any other known technique for detection of enzymatic activity. The detecting compound may be immobilized by any immobilizing agent e.g. agarose, agar, 15 gelatine, polyacrylamide, starch, filter paper, cloth; or any combination of immobilizing agents. For example, a-amylase activity can be detected by Cibacron Red labelled amylopectin, which is immobilized in agarose, α-amylase activity on this substrate produces zones on the plate with reduced red color intensity.

To screen for variants with increased stability, the filter with bound maltogenic 20 alpha-amylase variants can be pretreated prior to the detection step described above to inactivate variants that do not have improved stability relative to the parent maltogenic alpha-amylase. This inactivation step may consist of, but is not limited to, incubation at elevated temperatures in the presence of a buffered solution at any pH from pH 2 to 12, and/or in a buffer containing another compound known or thought to contribute to 25 altered stability e.g., surfactants, EDTA, EGTA, wheat flour components, or any other relevant additives. Filters so treated for a specified time are then rinsed briefly in deionized water and placed on plates for activity detection as described above. The conditions are chosen such that stabilized variants show increased enzymatic activity relative to the parent after incubation on the detection media.

To screen for variants with altered thermostability, filters with bound variants are incubated in buffer at a given pH (e.g., in the range from pH 2-12) at an elevated temperature (e.g., in the range from 50°-110°C) for a time period (e.g., from 1-20 minutes) to inactivate nearly all of the parent maltogenic alpha-amylase, rinsed in water, then placed directly on a detection plate containing immobilized Cibacron Red 35 labelled amylopectin and incubated until activity is detectable. Similarly, pH dependent stability can be screened for by adjusting the pH of the buffer in the above inactivation step such that the parent maltogenic alpha-amylase is inactivated, thereby allowing detection of only those variants with increased stability at the pH in question. To screen

for variants with increased calcium-dependent stability calcium chelators, such as ethylene glycol-bis(ß-aminoethyl ether) N,N,N',N'-tetraacetic acid (EGTA), is added to the inactivation buffer at a concentration such that the parent maltogenic alpha-amylase is inactivated under conditions further defined, such as buffer pH, temperature or a 5 specified length of incubation.

The variants of the invention may be suitably tested by assaying the starchdegrading activity of the variant, for instance by growing host cells transformed with a DNA sequence encoding a variant on a starch-containing agarose plate and identifying starch-degrading host cells as described above. Further testing in regard to altered 10 properties, including specific activity, substrate specificity, cleavage pattern, thermoactivation, thermostability, pH dependent activity or optimum, pH dependent stability, temperature dependent activity or optimum, transglycosylation activity, stability, and any other parameter of interest, may be performed on purified variants in accordance with methods known in the art as described below.

15 Degradation of β-limit dextrin by maltogenic alpha-amylase:

Another important parameter in the evaluation of the substrate specificity of maltogenic alpha-amylase variants may be the degree to which such enzymes are capable of degrading starch that has been exhaustively treated with the exoglycosylase B-amylase. To screen for variants which show patterns of degradation on such a 20 substrate differing from the patterns produced by the parent maltogenic alpha-amylase the following assay is performed: β-limit dextrin is prepared by incubating 25 ml 1% amylopectin in McIlvane buffer (48.5 mM citrate and 193 mM sodium phosphate pH 5.0) with 24 μg/ml β-amylase overnight at 30°C. Unhydrolysed amylopectin (i.e., β-limit dextrin) is precipitated with 1 volume 98% ethanol, washed and redissolved in water. 1 25 ml β-limit dextrin is incubated with 18 μl enzymes (at 2.2 mg/ml) and 100 μl 0.2 M citrate-phosphate pH 5.0 for 2 hrs at 30°C and analysed by HPLC as described above. Total hydrolysis of β-limit dextrin is carried out in 2M HCl at 95°C. The concentration of reducing ends is measured by methods known in the art.

Calcium binding affinity

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Unfolding of maltogenic alpha-amylases by exposure to heat or to denaturants such as guanidine hydrochloride is accompanied by a decrease in fluorescence, and oss of calcium ions leads to unfolding. Thus, the affinity of a maltogenic alpha-amylase variant for calcium can be measured by fluorescence measurements before and after incubation of the variant (e.g., at a concentration of 10 mg/ml) in a buffer (e.g., 50 mM 35 HEPES, pH 7) with different concentrations of calcium (e.g., in the range from

1 mM-100 mM) or of EGTA (e.g., in the range from 1-1000 mM) for a sufficiently long period of time (such as 22 hours at 55°C).

The measured fluorescence, *F*, is composed of contributions form the folded and unfolded forms of the enzyme. The following equation can be derived to describe the dependence of *F* on calcium concentration ([Ca]):

$$F = [Ca]/(K_{diss} + [Ca])(a_N - b_N log([Ca])) + K_{diss}/(K_{diss} + [Ca])(a_U - b_U log([Ca]))$$

where a_N is the fluorescence of the native (folded) form of the enzyme, b_N is the linear dependence of a_N on the logarithm of the calcium concentration (as observed experimentally), a_U is the fluorescence of the unfolded form and b_U is the linear dependence of a_U on the logarithm of the calcium concentration. K_{diss} is the apparent calcium binding constant for an equilibrium process as follows:

 K_{diss}

 $N-Ca \ll U + Ca(N = native enzyme; U = unfolded enzyme)$

In fact, unfolding proceeds extremely slowly and is irreversible. The rate of unfolding is dependent on calcium concentration, and such dependency for a given enzyme provides a measure of the calcium binding affinity of the enzyme. By defining a standard set of reaction conditions (e.g., 22 hours at 55° C), a meaningful comparison of $\mathcal{K}_{\text{diss}}$ for different maltogenic alpha-amylase variants can be made.

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Industrial Applications

The maltogenic alpha-amylase variants of the invention possesses valuable properties which may be advantageously used in various industrial applications. In particular, the enzyme finds potential application for retarding or preventing retrogradation, and thus the staling, of starch based food such as common in the baking industry.

The variant may be used for the preparation of bread and other bread products in accordance with conventional techniques known in the art.

It is believed that the modification of the starch fraction by use of the present invention results in increased volume in baked products and improved organoleptic qualities, such as flavour, mouth feel, palatability, aroma and crust colour.

The maltogenic alpha-amylase variant may be used as the only enzyme or as a major enzymatic activity in combination with one or more additional enzymes, such as xylanase, lipase, glucose oxidase and other oxidoreductases, or an amylolytic enzyme.

The enzyme variants of the invention also find industrial applicability as a component in washing, dishwashing and hard-surface cleaning detergent compositions. Some variants are particularly useful in a process for the manufacture of

linear oligosaccharides, or in the production of sweeteners and ethanol from starch, and/or for textile desizing. Conditions for conventional starch conversion processes, including starch liquefaction and/or saccharification processes, are described in, e.g., US 3,912,590 and in EP patent publications Nos. 252,730 and 63,909.

The invention is further illustrated with reference to the following examples which are not intended to be in any way limiting to the scope of the invention as claimed.

Determination of maltogenic amylase in MANU

One Maltogenic Amylase Novo Unit (MANU) is the amount of enzyme which under standard will cleave one µmol maltotriose per minute. The standard conditions are 10 mg/ml maltotriose, 37°C, pH 5.0, 30 minutes reaction time.

The pH dependence is found by repeating this measurement at the same conditions, but at different pH values.

EXAMPLES

15 Example 1: Construction of a variant of Novamyl with altered pH dependent activity

Novamyl is expressed in *Bacillus subtilis* from a plasmid denoted herein as pLBei010. This plasmid contains *amyM* in which the expression of *amyM* is directed by its own promoter and the complete gene encoding Novamyl, e.g., as contained in the strain DSM 11837. The plasmid contains the origin of replication, *ori*, from plasmid pUB110 and an kanamycin resistance marker for selection purposes. pLBei010 is shown in Fig. 1.

Primer sequences

Site directed mutants of Novamyl were constructed by the megaprimer method essentially as described by Kammann et al. (1989). Briefly, a mutagenic oligonucleotide primer is used together in a PCR reaction with a suitable opposite DNA strand end primer to create a preliminary PCR product. This product is then used as a megaprimer together with another opposite DNA strand end primer to create a double-stranded DNA product. The product of the final PCR reaction was routinely used to replace a corresponding DNA fragment in the pLBei010 plasmid by standard cloning procedures. Mutants were transformed directly into *Bacillus subtilis* strain SHa273, a derivative of *Bacillus subtilis* 168 which is apr, npr, amyE-, amyR2- and prepared by methods known in the art.

Oligonucleotide primers used in the construction of described variants are as listed below:

Variant Sequence (5'→ 3')

F188H: SEQ ID NO: 3

F188E: SEQ ID NO: 4

F284E: SEQ ID NO: 5

F284D: SEQ ID NO: 6

F284K: SEQ ID NO: 7

N327D: SEQ ID NO: 8

Variant Sequence (3'→ 5') 10

T288K: SEQ ID NO: 9

T288R: SEQ ID NO: 10

Aspartate variants of F284, T288 and N327 were obtained using primer A189 (SEQ ID NO: 11) and B649 (SEQ ID NO: 12) as end-primers.

F188-variants F188L, T189Y were obtained using primer A82 (SEQ ID NO: 13) 15 and B346 (SEQ ID NO: 14) as end-primers.

PCR products with the desired modification(s) were purified, digested with appropriate enzymes, separated by agarose gel electrophoresis and extracted, ethanol precipitated in the presence of glycogen, resuspended in H2O, ligated to pLBei010 20 which had been digested with the same appropriate enzymes, and transformed into Bacillus subtilis SHa273. Transformants were checked for size by colony PCR and for the insertion or removal of specific restriction sites by restriction enzyme digestion. Positive colonies were verified by DNA sequencing methods as described in the art.

Fermentation

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The B. subtilis SHa273 mutant clones were grown overnight on LB-Kana (10 μg/ml)-Starch plates at 37°C. The colonies from the plate were resuspended in 10 ml Luria broth. One-sixth of each of the suspensions were inoculated into a 500 ml shake flasks containing 100 ml PS-1 media, a soy meal/sucrose-based media, kanamycin for a final concentration of 10 $\mu g/ml$ and 100 μl 5M NaOH. The pH was adjusted to 7.5 with 30 NaOH before inoculation. The cultures were incubated for five days at 30°C with shaking at 270-300 rpm.

Enzyme Purification

Large particles from the media were removed by flocculation before affinity chromatography. Superfloc C521 (American Cyanmide Company) was used as the 35 cationic flocculant and Superfloc A130 (American Cyanmide Company) as the anionic flocculant.

The culture suspension was diluted 1:1 with deionized water and the pH was adjusted to approx. 7.5. A volume of 0.01 ml of 50^w/_w% CaCl₂ per ml diluted culture was added during stirring. A volume of 0.015 ml of 20^w/_w% Na-aluminate per ml diluted culture was titrated with 20% formic acid, while keeping the pH between 7 and 8. While stirring 0.025 ml 10^v/_v% of C521 per ml diluted culture was added, followed by 0.05 ml 1^w/_v% A130 per ml diluted culture, or until flocculation was observed. The solution was centrifuged at 4500 rpm for 30 minutes. Filtration was performed using a filter of pore size of 0.45 µm to exclude larger particles and any remaining bacteria. The filtered solution was stored at -20°C.

10 Immobilization of α-cyclodextrin to DSV-agarose

One hundred mg of α-cylcodextrin of molecular weight 972.86g/mol (Fluka 28705) was dissolved in 20 ml coupling buffer (0.5M Na₂CO₃, pH 11). Ten ml of DSV-agarose (Mini-Leak, Medium 10-20 mmol/l of divinyl sulfone activated agarose (Kem-En-Tec) was washed thoroughly with deionized water, then dried by suction and transferred to the a-cyclodextrin solution. After the mixture had stirred for 24 hr at ambient temperature, the gel was washed with deionized water, followed by 0.5M KHCO₃. The gel was transferred to the blocking buffer (20ml 0.5M KHCO₃ + 1ml mercaptoethanol), stirred for 2 hr at ambient temperature, then washed with deionized water.

20 Affinity chromatography

The variants were purified by affinity chromatography using the Pharmacia FPLC System. A 0.04 volume of 1M Na-acetate pH 5 was added to the filtrate obtained by flocculation to adjust pH and CaCl₂ was added to a final concentration of 10⁻¹⁰ M. The solution was filtered and degassed. A Pharmacia XK16 column was prepared with ten ml of the immobilised α-cyclodextrin, then equilibrated in the equilibration buffer (25 mM Na-acetate pH 5) by washing with approximately 10 times the column volume. The filtrate was applied to the XK16 column, which was then washed with the equilibration buffer until protein could no longer be detected in the washing buffer. The column was washed with the equilibration buffer containing 0.5M NaCl to elute nonspecific material, followed by another wash with 2-3 times the column volume of the equilibration buffer. All washings were performed using a flow rate of 10ml/min. Specifically bound material was eluted using a solution of 2% α-cyclodextrin in the wash buffer and collected using the Pharmacia Liquid Chromatography Collector LCC-500 Plus using a flow rate of 5 ml/min.

Example 2: pH dependent activity of variants

The variants prepared in the preceding Example were tested for activity at various pH values as follows.

A colorimetric glucose oxidase-peroxidase assay for liberated glucose from maltotriose or amylopectin was used to determine the pH activity profiles of the enzyme variants (Glucose/GOD-Perid® Method, Boehringer Mannheim, Indianapolis IN). Activity was assayed in a buffer of 25 mM citrate-phosphate, 0.1mM CaCl₂ at pH values of 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8 and 8.6. The buffer pH was adjusted using NaOH and enzymes were diluted in 25 mM citrate-phosphate buffer pH 5. Measurements were taken in duplicate to obtain an average value. All values are relative to the pH at which the highest level of activity is seen.

The results, shown in the table below, indicate that each of the variants has an alteration in the pH dependent activity profile when compared to the parent Novamyl[®]. The highest level of activity for each variant is designated 100% and the activity of that variant measured at the other indicated pH values is a relative percentage of that maximum.

Modifications		рН												
	2.0	2.5	3.0	3.5	4.0	4.5	5.0	5.5	6.0	6.5	7.0	7.5	8.0	8.6
None (parent)	0	0	0	8	47	80	100	95	91	80	66	39	35	30
F188H	1	0	0	1	3	29	77	99	100	88	59	39	31	27
F188E	0	0	0	2	27	62	89	100	93	71	46	28	20	18
T288R	0	0	0	8	51	7 7	94	100	86	73	50	34	27	12
N327D	1	1	7	27	67	95	100	98	77	33	19	11	5	0

Further, a number of Novamyl variants were tested for activity at pH 4.0 and 5.0, taking the activity of Novamyl at the same pH as 100 %. The activity was determined by hydrolysis of maltotriose (10 mg/ml) at 60°C, 50 mM sodium acetate, 1 mM CaCl₂. The results are expressed as the ratio between activity at pH 5.0 and pH 4.0:

Modifications	pH 5.0/pH 4.0		
N131D	0.24		
I174Q	0.31		

G397P	0.40
H103Y	0.40
△ 262-266	0.47
S32Q	0.53
\$32D	0.55
T142A+ D261G	0.62
G370N+ N371G	0.66
S32N	0.68
N176S	0.79
D17E	0.80
None (parent)	1
∆ 191	1.39
192-A-193	1.61
1174E	1.80
192-A-G-193	1.90
△ 192	2.22

The results demonstrate that variants with a higher or lower pH optimum can be obtained according to the invention.

Example 3: Thermostability of variants

5 Incubation at 80°C

The thermostability of a number of Novamyl variants was tested by incubating an aqueous solution at 80°C and pH 4.3 and measuring the residual amylase activity at various times. The parent enzyme, Novamyl, was included for comparison. The results are expressed as residual activity at various times in percent of initial activity:

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Modifications	0	5 min.	10 min.	15 min.	20 min.	25 min.
None (parent)	100	23	9	3	1	0
F188L+ V336L+ T525A	100	63	49	48	52	47
F188I+ Y422F+ I660V	100	71	6 0	51	43	38
N115D+ F188L	100	73	60	51	44	39

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A30D+ K40R+ D261G	100	38	24	15	13	10
T142A+ N327S+ K425E+ K520R+ N595I	100	47	39	25	19	11
F188L+ D261G+ T288P	100	60	67	66	63	67
K40R+ F188L+ D261G+ A483T	100	56	48	40	36	30
T288K	100	64	31	18	7	4

The above data show a clearly improved thermostability for the variants compared to the parent amylase.

Incubation at 85°C with calcium

The Novamyl variant S32E was tested by incubation with 1 mM Ca⁺⁺ at 85°C for 15 minutes. The variant showed a residual activity of 48 % whereas the parent enzyme (Novamyl) showed 32 % residual activity at the same conditions.

DSC

Further, the thermostability was tested for some Novamyl variants by DSC (differential scanning calorimetry) at pH 4.3 or 5.5. Again, the parent amylase was included for comparison. The results are expressed as the denaturation temperature (Tm):

Modifications	Tm at pH 4.3	Tm at pH 5.5
None (parent)	79°C	88°C
N115D+ F188L	86°C	92°C
T142A+ N327S+ K425E+ K520R+ N595I	not determined	93°C

The results show improved thermostability for both variants.

Example 4: Specific activity of variants

Amylase activity was determined by a colorimetric measurement after action on Phadebas tablets at pH 5.0 and 60°C. The results for two Novamyl variants, relative to Novamyl were as follows:

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Modifications	Relative amylase activity
None (parent)	100
192-A-193	110
△ (191-195)	300

The specific activity was further tested by action on maltotriose at pH 4.0, 60°C by the MANU method described above. The results showed that the variant G370N, N371G has a maltotriose activity of 106 % compared to Novamyl.

5 Example 5: Inhibition of retrogradation

The efficiency of Novamyl and Novamyl variants to inhibit retrogradation was determined as follows:

730 mg of 50 % (w/w) amylopectin slurry in 0.1 M sodium acetate, at a selected pH (3.7, 4.3 or 5.5) was mixed with 20 µl of an enzyme sample, and the mixture was incubated in a sealed ampoule for 1 hour at 40°C, followed by incubation at 100 °C for 1 hour in order to gelatinize the samples. The sample was then aged for 7 days at room temperature to allow recrystallization of the amylopectin. A control without enzyme was included.

After aging, DSC was performed on the sample by scanning from 5°C to 95°C at a constant scan rate of 90°C/hour. The area under the first endothermic peak in the thermogram was taken to represent the amount of retrograded amylopectin, and the relative inhibition of retrogradation was taken as the area reduction (in %) relative to the control without enzyme.

In the table below, the efficiency of the enzyme is expressed as the ratio of the relative inhibition of retrogradation to the enzyme dosage (in MANU/ml):

рН	Modifications	MANU/ml	Relative inhibition	Efficiency
3.7	A30D+ K40R+ D261G	0.23	0.38	1.7
3.7	T142A+ N327S+ K425E+ K520R+ N595I	0.07	0.29	4.1
3.7	None (parent)	0.27	0.38	1.4
4.3	N115D+ F188L	0.01	0.18	18
4.3	None (parent)	0.27	0.43	1.6

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5.5	∆ (191-195)+ F188L+ T189Y	0.02	0.12	6
5.5	∆ (191-195)	0.02	0.14	7
5.5	∆ (191-195)	0.05	0.31	6.2
5.5	N115D+ F188L	0.01	0.39	39
5.5	T142A+ D261G	0.14	0.53	3.8
5.5	None (parent)	0.27	0.49	1.8

The results demonstrate that a number of variants are more efficient than the parent amylase to inhibit retrogradation.

Example 6: Anti-staling effect of variants

Bread was made by the European Straight Dough method or from sour dough with or without addition of enzymes, and loaves were baked in lidded pans, to avoid volume effects. The bread was allowed to cool for 2 hours, and the texture was analyzed. The remaining loaves were then wrapped in plastic bags and stored at room temperature for texture analysis after 1, 4 and 7 days.

The texture analysis of each loaf was done by cutting 4 slices; the force was measured at 25 % compression (P1), at 40% compression (P2) and after keeping 40% compression constant for 30 sec. (P3). P1 was taken as the firmness, and the ratio (P3/P2) was taken as the elasticity of the crumb. The extent of retrogradation after 7 days storage was determined by DSC as described in Example 7.

15 European Straight Dough (pH5.5-6.0)

A Novamyl variant (T142A+ N327S+ K425E+ K520R+ N595I) was tested at dosages in the range of 0-2 mg enzyme/kg flour, and the parent enzyme (Novamyl) was used for comparison.

The results showed that at equal dosages, the variant gives better elasticity (P3/P2) than the parent enzyme after two hours and 1 day. The results after 7 days showed that the variant at dosages of 1-2 mg/kg gave significantly softer crumb (lower firmness, P1) than the parent enzyme at the same dosage. Thus, the variant has a better anti-staling effect throughout a 7-day storage period.

Sour dough (pH approx. 4.5)

A Novamyl variant (F188L+ D261G+ T288P) was tested in sour dough, and the parent enzyme (Novamyl) was used for comparison. The following results were



obtained for firmness (P1) after 7 days, elasticity (P3/P2) after 4 and 7 days and ret-

rogradation after 7 days:

Enzyme	Dosage mg/kg flour	Firmness (P1) after 7 days
None	0	2590
	1	2031
Parent	3	1912
	13	1570
Variant	1	1436
	3	1226

Enzyme	Dosage mg/kg flour	Elasticity 4 days	Elasticity 7 days
None	0	0.49	0.47
	1	0.51	0.52
Parent	3	0.53	0.51
	13	0.53	0.51
Variant	1	0.59	0.57
	3	0.57	0.58

Enzyme	Dosage mg/kg flour	Retrogradation, 7 days (relative to control)
None	0	100 %
	1	100 %
Parent	3	63 %
	13	32 %
Variant	1	46 %
	3	20 %

The results show that the variant has a markedly improved effect on texture evaluated as firmness and elasticity in sour dough at pH 4.5. A dosage of 1-3 mg/kg of the variant is superior to 13 mg/kg of the parent enzyme on all parameters tested, and the elasticity achieved with the variant cannot be matched by the parent enzyme at any dosage.

Example 7: Cleavage pattern of variants

The cleavage pattern in starch hydrolysis was compared for two variants and the parent enzyme, Novamyl.

The results below indicate % by weight of each oligosacccharide (G1-G8) formed after 24 hours incubation in 1% (w/v) starch using 50 mM sodium acetate, 1 mM CaCl₂, pH 5.0 at 50 °C. The oligosaccharides were identified and quantitated using HPLC.

Oligosaccharide	Parent	△ (191-195)	N115D+ F188L
G8	-	1.7	-
G 7	-	2.6	-
G6	-	7.5	1.4
G 5	-	10.1	2.1
G4	-	21.1	11.3
G 3	-	28.7	10.7
G2	96.5	28.3	61.9
G1	3.5	-	12.6

The results demonstrate a significantly altered cleavage pattern. Novamyl after 24 hours produces mainly maltose and virtually no higher oligosaccharides. In contrast, the two variants produce significant amounts of maltotriose and higher oligosaccharides.

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70.305	68.798	69.311	70.684	69.618	71.409	67.560	66.675	66.234	66.203	65.392	65.699	06.240	66.545	62.609	66.206	66.444	65.787	65.261	63.801	63.527	65.607	65.084	65.851	62.881	61.460	60 495	60.894	59.363	58.741	58.722	61.857	61.698	62.727	62.836	63.414	64.406
27.188	25.298	24.907	25.345	25.067	23.436	25.047	24.178	23.007	23.116	24.945	26.104	21.215	28.295	27.004	28.139	29.227	21.932	20.774	20.582	20.020	18.280	18.376	17.115	21.153	21.160	21.628	19.763	19.772	20.803	18.647	22.797	23.494	23.096	23.819	21.975	21.535
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79.475	79.868	81.181	81.716	82.088	83.383	78.307	79.126	79.182	80.101	78.421	78.508	79.662	77.488	277.67	77.621	78.740	78.220	78.329	78.622	78.215	77.031	75.766	75.896	74.716	79.460	79.667	80.939	81.252	79.785	79.524	78.624	80.201	81.725	83.019	82.901	83.825	83.609	81.987	81.973	81.014
31.171	30.414	30.576	32.069	31.387	31.377	29.904	30.073	28.855	28.803	31.228	32.504	33.310	32.804	34.466	33.940	34.769	27.917	26.731	25.486	24.375	26.557	26.500	26.191	26.767	25.656	24.557	23.787	22.840	25.253	24.264	23.408	24.417	24.109	23.428	21.914	21.341	23.828	21.317	19.845	19.237
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65.781	56.264	63.334	65.711	66.280	67.573	68.698	68.912	67.984	69.243	69.277	70.381	70.407	71.530	71.515	72.601	69.456	70.658	71.939	72.876	70.683	69.510	69.391	70.788	72.003	73.257	73.337	72.513	74.369	74.586	76.085	76.850	73.957	74.647	75.727	74.060	76.484	77.924	78.827	80.029	78.072
22.181	22.048	19.488	19.569	22.977	23.613	22.652	21.639	24.440	25.238	26.324	24.989	27.139	25.773	26.843	27.673	22.895	22.120	22.877	22.290	21.886	21.045	20.812	19.828	24.138	24.888	26.169	26.402	26.981	28.205	28.353	27.377	29.438	29.783	29.311	30.650	29.559	29.714	29.665	29.444	31.055
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87.603	89.341	90.225	91.232	87.768	86.771	86.833	85.746	87.051	86.796	85.340	84.389	000.00	82.597	87.990	87.980	87.277	86.403	89.446	89.417	89.009	89.736	88.936	89.265	89.156	87.528	86.836	85.311	84.533		88.769	89.046	88.229	90.199	84.945	83.521	82.709	81.542
30.292	31.997	30.831	30.462	29.115	28.057	27.585	27.261	26.893	27.286	27.797	26.694	00.02	25.769	27.508	27.104	28.195	27.826	26.915	26.645	25.446	27.712	25.242	26.275	26.097	29.473	30.468	30.442	30.482	31.939	32.131	33.640	34.487	34.009	30.314	30.436	29.302	29.555
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24.483	5.269	660.80	29.205	29.219	28.461	7.206	27.692	26.282	30.136	31.258	30.770	31.500	32.359	32.995	33.210	33.408	33.830	34.021	34.208	34.816	29.675	29.112	28.587	28.651	27.940	28.518	27.397	26.462	27.387	28.162	7.702	28.836	28.746	26.793	5.528	1.749	3.657		5.816
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70.400	72.780	65.76	65.200	63.996	65.961	66.28	66.701	65.052	66.037	62.53	65.090	64.306	66.586	66.907	65.941	68.228	66.245	68.569	67.56	67.856	65.62	65.088	63.69	62.875	66.001	67.35	68.22	67.699	69.518	63.349	61.979	61.000	59.868	61.994	62.793	62.165	63.06	62.37	62.030
9.398	582	730	11.936	11.665	14.221	091	16.493	15.227	11.480	9/9	9.294	674	285	11.928	882	163	14.103	379	14.319	15.536	169	501	677	712	910	530	910	355	6.372	881	163	328	8.839	10.397	116	958	8.449	682	414
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75.523	74.684	76.	76.	77.	77.	78.	75.	74.	73.	72.	75.	75.	72.	71.	70.	69	71.	70.170	69	70.350	71.	70.	69	68	70.889	70.	70.	69.448	68.474	71.	71.	71.113	72.178	69	68.449	67.	99	68	70.
16.053	712	259	889	577	131	197	600	611	900	160	260	488	300	726	420	924	255	816	202	333	710	427	923	565	910	062	609	365	989	930	507	899	223	825	523	166	349	232	924
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45.468	44.545	43.692	42.710	44.043	45.128	46.061	45.505	46.238	46.481	47.790	49.023	47.999	44.168	43.586	43.872	44.184	42.039	41.392	40.654	41.597	43.719	43.907	45.292	45.486	43.745	42.397	43.984	46.268	47.655	47.985	48.642	48.605	50.051	48.191	47.430	47.642	47.144	47.806	46.851	47.591	46.690
75.092	76.068	77.042	77.631	77.214	73.464	72.569	71.940	71.757	71.419	70.697	71.580	69.510	71.683	71.091	71.937	71.401	71.000	70.507	71.318	69.245	73.275	74.106	73.906	73.741	75.596	75.753	76.459	73.847	73.732	72.341	72.184	74.237	74.258	75.632	71.287	69.911	69.830	69.251	68.979	68.833	67.579
33.806	30.773	29.979	30.499	28.773	32.248	32.959	34.219	35.179	31.968	32.367	32.357	31.384	34.226	35.441	36.687	37.751	35.269	36.571	37.171	36.866	36.588	37.769	38.367	39.593	37.365	36.906	38.641	37.440	37.915	38.401	39.438	36.780	37.297	36.323	37.764	38.262	39.715	40.554	37.342	36.004	38.025
ASP A	ASP A	ASP A	ASP A	ASP A	LEU A	LEU A	LEU A	LEU A	LEU A	LEU A	LEU A	LEU A	ASN A	ASN A	ASN A	ASN A	ASN A	ASN A	ASN A	ASN A	THR A	THR A	THR A	THR A	THR A	THR A	THR A	VAL A 289	VAL A	VAL A	VAL A	VAL A	VAL A	VAL A	ILE A	ILE A	ILE A	ILE A	ILE A	ILE A	ILE A
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2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275	2276	2277	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306
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36.365	38.864	39.263	39.990	38.787	38.492	37.977	38.660	39.799	38.187	37.397	39.638	37.904	38.460	39.476	39.298	37.283	36.538	35.102	37.196	40.581	41.764	41.696	42.316	43.014	44.271	44.259	45.290	40.834	40.731	41.966	42.313	39.517	38.227	37.520	37.765	36.386	36.596	35.903	42.687	43.882	44.885
71.116	69.368	69.056	69.805	67.913	71.312	71.621	70.782	70.318	73.104	74.024	73.569	70.675	70.087	71.063	72.295	69.883	68.558	68.726	67.351	70.458	71.304	71.778	71.275	70.451	71.343	72.526	70.758	72.815	73.424	74.271	74.331	74.409	73.575	72.991	73.416	72.232	72.665	72.085	74.753	75.575	74.670
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46.990	48.549	48.874	46.812	44.770	49.472	50.881	50.984	51.952	51.610	52.199	51.385	53.565	51.854	54.064	53.231	50.183	50.270	50.925	51.220	48.870	48.035	48.250	51.194	51.975	53.114	53.094	51.214	50.299	49.668	48.827	50.107	54.106	55.252	55.355	54.644	56.601	56.804	56.464	56.443
70.499	71.894	72.436	73.276	72.308	71.647	72.026	73.543	74.035	71.271	69.924	68.917	69.711	67.656	68.444	67.450	74.322	75.773	76.303	77.505	76.437	75.982	76.059	75.400	75.785	74.738	73.753	75.808	77.072	77.320	76.525	78.376	75.020	74.106	73.485	73.874	74.850	75.771	75.587	72.672
46.271	47.675	48.755	46.005	46.976	46.760	46.917	47.104	47.719	48.052	47.630	47.153	47.760	46.821	47.438	46.948	46.411	46.398	45.131	45.018	46.488	45.446	47.869	44.162	43.009	42.852	43.624	41.654	41.692	40.301	39.959	39.635	42.031	41.924	40.518	39.580	42.152	41.116	43.511	40.337
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2349	2351	2352	2354	2355	2356	2357	2358	2359	2360	2361	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390
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46.700	45.455	46.293	45.380	43.110	41.647	40.907	40.109	40.068	39.363	46.801	47.653	49.051	49.634	47.750	46.379	45.665	46.267	49.542	50.940	50.969	51.783	51.499	52.820	51.736	50.033	49.960	48.964	49.093	49.553	50.584	51.808	50.295	52.724	51.232	52.463	47.976	46.907	47.187	47.612
68.108 70.386	70.246	71.071	70.770	69.717	70.173	70.343	69.361	68.126	69.593	72.204	73.001	72.376	72.565	74.471	75.238	74.831	76.242	71.574	71.075	69.743	69.520	70.961	70.175	72.364	68.828	67.537	67.464	66.540	66.455	66.334	65.683	66.865	65.579	66.762	66.104	68.355	68.234	69.187	68.676
34.964	41.352	42.343	43.481	40.602	40.480	41.729	42.225	41.706	43.322	41.972	42.894	42.983	44.074	42.408	42.593	43.466	41.735	42.009	42.157	42.861	43.748	40.703	40.707	40.153	42.479	43.106	44.255	45.095	42.063	40.936	41.178				38.880			46.504	47.547
2307 CD1 ILE A 290 2308 N ARG A 291	CA ARG A	C ARG A	CB ARG A	CG ARG A	CD ARG A	NE ARG A	CZ ARG A	NH1 ARG A	NH2 ARG A	N ASN A	CA ASN A	C ASN A	O ASN A	CB ASN A	CG ASN A	OD1 ASN A	ND2 ASN A	N VAL A	CA VAL A	C VAL A	O VAL A	CB VAL A	CG1 VAL A	CG2 VAL A	N PHE A	CA PHE A	C PHE A	O PHE A	CB PHE A	CG PHE A	CD1 PHE A	CD2 PHE A	CE1 PHE A	CE2 PHE A	CZ PHE A	N GLY A	CA GLY A	C GLY A	O GLY A
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58.967	54.848	53.499	53.225	56.143		56.155	51.254	51.272	50.512	50.252	48.767	48.246	48.324	52.340	52.822	52.321	53.406	53.770	52.727	54.264	53.176	53.030	55.470	56.684	57.595	52.426	51.422	50.319	49.942	50.869	49.859	49.819	50.019	49.600
74.651		76.830	77.409	78.686	78.499	79.477	75.915	74.957	75.156	75.431	75.462	77.075	78.010	73.136	74.049	73.974	72.052	71.356	71.038	75.848	76.824	77.017	76.680	74.725	76.208	77.423	78.422		78.286	79.123	80.214	81.169	80.762	82.401 76.630
31.910	32.418	31.740	30.672	33.863	33.275	34.959	31.750	30.590	29.639	32.849	32.375	31.759	33.282	29.383	28.272	27.153	29.712	28.388	30.641	27.479	27.073	25.875	27.997	28.109	28.967	27.970	27.549	26.734	25.672	28.821	28.507	29.734	30.875	29.417
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4.325			64.127		62.651	65.680	65.987		65.102	65.323	63.807	63.207	63.200	65.749	65.359	65.556		66.162	67.636				65.393		66.836	64.740	65.577			66.668		67.808	69.155		71.012		69.815	69.188	243	vo	69.433
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37.288	39.550	38.139	37.386	39.60	37.571	38.05	38.76	37.81	36.678	40.149	40.16	39.080	41.23	38.279	37.486	38.33	39.57	36.17	36.35	37.35	35.33	37.80	38.55	39.03	39.74	37.72	36.51	35.652	36.01	34.648	34.85	38.746	39.181	40.07	40.38	37.955	37.069	37.477	35.891	40.56	41.53
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13.17	0.64	10.68	10.60	10.70	11.96	9.53	9.68	13.05	9.85	9.04	9.68	11.13	10.40	10.10	9.96	2.80	9.02	9.87	8.82	9.73	10.64	0.77	0.73	90.0	9.44	10.80	9.58	10.73	9.57	9.05	11.94	11.07	11.10	11.33	13.68	9.28	9.24	6.67	10.76	1.75	10.90
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66.275	67.901	68.499	68.918	67.643	66.685	69.733	71.176	71.820	72.064	67.671	66.478	66.821	67.716	66.118	65.842	64.954	64.698	66.133	66.524	65.704	64.628	66.302	64.919	67.237	66.234	65.582	66.205	67.375	65.698	67.122	67.817	67.746	69.142	69.083	69.788	65.385	65.879	66.277	67.116	64.869	63.705
18.619	. 558	23.285	24.610	25.415	25.448	.299	24.761	25.310	25.183	.185	27.014	.477	29.071	.872	25.387	27.793	.773	.019	30.333	31.433	31.218	30.364	30.009	29.314	32.660	33.784	.086	35.086	.716	.638	32.421	. 798	32.321	34.683	.498	6.137	37.469	38.277	9.255	18.211	677.
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0.144	39.639	40.421	40.438	38.179	7.436	41.169	42.094	3.392	3.962	12.447	3.441	1.154	43.852	5.132	44.976	44.865	6.272	7.621	47.738	48.694	44.939	4.747	45.993	5.858	44.225	45.154	7.095	48.297	48.193	8.134	49.485	50.787	51.110	51.502	48.148	47.941	9.122	906.8	7.665	6.279	46.055
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.014	19.117	.115	.159	8.843	.221	8.041	1.092	1.805	.593	169.91	.646	0.6.	1.525	.114	.153	3.082	1.141	3.570	49.572	.865	.364	1.477	.257	.294	. 884	.871	9.847	.601	18.267	.246	.554	.516	.403	.648	48.283	.008	6.077	14.859	.345	900.8	.318
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9.51	1.47	3.17	1.23	5.96	3.78	10.04	9.86	1.24	1.71	11.21	14.33	11.49	11.49	11.63	2.21	13.45	16.61	0.26	15.50	12.48	10.61	0.74	9.55	0.54	12.40	13.01	11.67	12.76	15.62	18.86	19.61	15.77	5.46	10.49	1.60	2.74	3.05	3.94	3.29	4.98	3.51
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330	67.156	015	973	743	206	680	618	557	892	207	981	321	64.272	.603	65.103	914	516	61.101	60.592	59.753	59.388	251	64.168	431	916	709	63.676	64.006	63.230	041	513	342	261	763	62.257	035	864	748	910		905
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2.537	42.224	0.858	10.005	19.087	17.923	13.702	4.650	4.130	13.295	600.91	15.623	14.540	14.057	14.499	15.591	14.667	13.997	13.560	13.017	11.965	11.546	11.440	13.654	4.019	5.461	16.170	13.007	3.365	4.315	2.718	4.651	3.014	3.978	5.825	7.183	18.303	9.456	17.269	16.461	16.488	17.045
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61.812	57.761	57.810	58.444	57.127	61.192	62.559	62.958	62.168	62.702	62.516	64.198	64.730	65.599	65.974	65.791	65.203	65.817	66.694	67.749	67.451	65.860	62.009	62.629	63.667	64.882	62.914	63.551	62.811	69.008	70.040	69.912	69.819	71.418	71.733	69.758	69.529	70.863	71.131	68.403	67.136
62.543	62.971	61.634	61.462	60.605	63.161	63.516	62.739	62.718	65.025	65.770	62.251	61.703	62.835	63.761	60.598	59.420	62.845	63.886	63.156	62.293	64.824	65.807	66.940	65.549	67.843	66.474	67.595	68.508	63.663	63.232	64.002	65.229	63.684	62.764	63.265	63.892	63.757	62.703	63.188	63.120
52.198	53.543	54.248	55.292	53.729	50.115	49.738	48.481	47.524	49.451	50.658	48.417	47.177	46.581	47.366	47.452	48.033	45.302	44.693	43.877	43.032	43.772	44.592	45.124	44.836	45.908	45.595	46.107	46.871	43.976	43.034	41.698	41.739	43.620	44.701	40.628	39.303	38.498	37.935	38.537	39.386
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58.922	56.120	55.689	54.758	58.596	58.943	57.715	57.309	60.036	60.383	60.026	61.224	57.181	55.958	56.267	55.310	55.161	55.869	55.043	55.738	55.195	53.948	55.947	57.513	57.734	57.869	58.830	59.071	59.255	58.985	59.700	56.949	56.883	57.846	57.917	55.456	54.990	54.554	58.501	59.481	60.910
64.734	62.608	63.352	•	62.933	63.646	63.653	62.621	62.821	63.480	64.602	62.721	64.891	64.887	64.725	64.552	66.201	67.415	68.719	69.847	71.046	71.299	72.058	65.035	65.048	63.623	62.900	65.787	66.167	65.335	67.376	63.224	61.822	61.585	60.450	61.431	62.330	61.611	63.669	62.467	62.729
39.366	36.749	39.146	36.230	40.741	41.963	42.852	43.367	42.711	44.030	44.422	44.789	43.055	43.878	45.362	46.117	43.673	44.296	44.031	44.772	44.957	44.521	45.406	45.779	47.255	47.780	47.440	47.474	48.921	49.775	49.263	48.650	49.087	50.271	50.750	49.518	50.542	48.284	50.666	51.778	51.339
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61.907	59.626	59.340	60.269	60.410	57.882	57.650	56.882	60.914	61.883	61.508	61.324		63.638	63.904	63.716	64.166	64.049	63.546	64.216	63.701	64.044	61.394	60.903	61.637	62.207	59.410	59.117	58.993	58.485	59.455	61.523	62.092	61.102	60.614	63.393			63.800	65.176		65.003
55.439	55.975	56.313	55.457	54.276	55.903	56.154	56.812	56.135	55.413	55.715	56.878	55.953	55.807	54.628	56.884	54.969	56.328	58.271	57.117	59.052	58.461	54.618	54.869	54.121	53.092	54.452	52.972	52.219	52.726	50.952	54.698		54.203	55.341	54.774				52.511	•	53.254
36.431	33.950	32.567	31.685	31.910	32.231	30.712	32.978	30.771	29.900	28.446	28.103	30.222	31.708	32.381	32.632	33.717	33.883	32.509	35.020	33.641	34.894	27.680	26.332	25.221	25.472	26.259	26.541	25.208	24.222	25.214	24.024	22.819	21.665	21.507	22.452	21.152	21.146	19.963	19.956	18.770	18.817
SER A	VAL A	VAL A	C VAL A 489	O VAL A 489	CB VAL A 489	VAL	CG2 VAL A 489	TRP		TRP A	O TRP A 490	TRP A	TRP A	4	TRP A	TRP A	CE2 TRP A 490	TRP A	TRP A	TRP A	TRP A	æ	GLN A	C GLN A 491	GLN A	GLN A	GIN A	GLN A	GLN A	2 GLN A	TYR A	_	C TYR A 492	TYR A	TYR A	CE2 TYR A 492	TYR A				
	•	3848		3850	3851	3852	3853	3854	3855	3856	3857	3858	3859	3860	3861	3862	3863	3864	3865	3866				3870	3871	3872	3873	3874	3875	3876	3877	3878	3879	3880	3881	3882	3883	3884	3885	3886	3887
9	7	9	9	œ	9	9	9	9	7	9	9	8	9	7	9	9	80	9	9	9	7	9	9	80	7	9	9	ω	9	7	9	9	œ	9	9	9	7	9	9	89	9
1.00 20.55	14	1.00 13.06	7	1.00 15.20	1.00 13.14	1.00 14.96		1.00 14.36	1.00 12.65	1.00 13.54	1.00 13.15	1.00 13.10	1.00 12.82	1.00 14.66	1.00 11.27	1.00 12.98	1.00 13.40	1.00 15.48	1.00 15.92	1.00 15.61	1.00 14.03	1.00 13.19	1.00 16.21	1.00 13.21	1.00 11.65	1.00 13.98	1.00 15.41			1.00 12.17	1.00 9.79	1.00 9.96	1.00 12.30	1.00 10.95	1.00 12.89	1.00 15.13	1.00 11.84		1.00 11.64	1.00 12.81	1.00 15.82
	66.454	64.991	64.394	64.487	64.486	65.33	62.720	62.727	63.680	63.175		61.387	62.782	61.687	60.632	59.301	58.884	60.545	61.936	62.333	58.671	57.309	57.376	56.271	58.553	58.545	58.068	58.196	60.002	57.647	57.403	58.257	58.179	55.919	55.762	54.964	59.078	59.865	59.693	59.650	61.336
58.894	59.860	59.690	59.527	60.432	60.917	60.936	59.717	62.242	58.414	58.090	58.961	59.723	56.581	58.980	59.827	59.496	58.331	59.546	58.945	58.160	60.548	60.352	59.600	59.235	59.349	58.395	58.969	60.148	57.937	58.008	58.220	57.174	55.970	58.022	58.150	58.990	57.635	56.623	56.972	58.154	56.705
44.953	42.216	42.004	43.383	44.199	41.231	40.812	39.938	40.073	43.621	44.961	45.370	44.562	44.892	46.637	47.183	46.509	46.374	48.704	48.959	47.698	45.977	45.399	44.067	43.561	43.508	42.375	41.058	40.730	42.233	40.213	38.776	38.057	38.406	38.428	36.871	39.127	37.112	36.335	34.867	34.519	36.850
CG2 THR A 481	I LEU A 482	CA LEU A 482	LEU A	LEU A	CB LEU A 482	LEU A	CD1 LEU A 482	LEU A	ALA A	A ALLA A	ALA	Ø	ALA	PRO A	PRO A	ø	PRO A	PRO A	PRO A	PRO A	GLY A		GLY A	GLY A	ALA A	ALA A	ALA A	ALA A	CB ALA A 486	VAL A	CA VAL A 487	VAL A	VAL A	3 VAL A	VAL A		SER A	CA SER A 488	SER A	O SER A 488	SER A
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3970 N	PRO A	11.674	64.766	38.661		7	4012	z	GLY A 512	0.462	65.892	33.876	1.00 12.55	7
ີວ	PRO A	12.353	66.020	38.410	11.	9	4013	ð	⋖	0.554	64.629	33.165	1.00 14.21	9
O	PRO A	11.747	67.182	39.194	1.00 12.55	v	4014	U	A	1.950		33.170		y
0	PRO A	10.711	67.015	39.862		œ	4015	0	ø	2.002	62.765	32.923		æ
បី	PRO A	12.227	66.313	36.887		9	4016	z	ASN A 513	2.994	64.789	33.454	1.00 12.18	7
ช	PRO A	11.545	65.090	36.310	1.00 15.76	9	4017	ర	Ø	4.306	64.154	33.561	10	9
บี	PRO A	11.007	64.329	37.471	1.00 14.19	9	4018	ပ	ø	4.360	63.144	34.731	15	9
z	ASN A	12.364	68.381	39.142	1.00 10.92	7	4019	0	Ø	3.699	63.392	35.738	13	80
Ü	ASN A	11.835	69.493	39.909		9	4020	ප	4	5.408	65.230	33,833	Ξ	9
	ASN A	10.940	70.447	39.087	1.00 10.54	9	4021	ន	ASN A 513	5.762	66.023	32.583		ø
	ASN A	10.497	71.416	39.701		80	4022	go	Ø	6.738	66.874	32.746	16	æ
	ASN A	13.043	70.353	40.401	1.00 11.79	9	4023	ND2	4	5.078	65.931	31.505	1.00 8.90	7
	ASN A	14.033	69.428	41.196	1.00 14.07	9	4024	z	K	5.280	62.177	34.576	Ξ	۲
	ASN A	15.192	69.361	40.778	1.00 13.37	60	4025	ర్	4	5.505	61.269	35.745		G
	ASN A	13.592	68.845		1.00 13.47	7	4026	O	Ø	6.594	61.894	36.612	7	9
	MET A	10.654	70.046		1.00 10.20	7	4027	0		7.617	62.379	36.131	1.00 15.35	80
	MET A	9.823	70.957			9	4028	9	VAL A 514	6.072	59.931	35.221		•
	MET A	9.102	70.087		1.00 11.63	9	4029	G	Ø	6.529	59.036	36.390	14	9
	MET A	9.633	69.059			œ	4030	CG2	Ø	5.042	59.176	34.393		9
	MET A	10.929	71.782			9	4031	z		6.335	61.959	37.923	1.00 10.41	7
	MET A	10.270	72.808			ø	4032	ర్	¥	7.256	62.496	38.927	12	9
	MET A	11.558	73.692			16	4033	ပ	A	7.433	61.442	40.001		9
បី	MET A	11.921	72.582	33.003		ø	4034	0	æ	6.495	60.711	40.306	1.00 14.78	1
	GLY A	7.935	70.545			7	4035	8	Ą	6.563	63.772	39.511	7	9
ರ	GLY A	7.253	69.739			v	4036	CG	ď	7.228	64.271	40.775	79	9
υ	GLY A	5.851	70.331			ø	4037	CG2	K	6.678	•	38.435	1.00 16.90	o
0	GLY A	5.506	71.321	34.	1.00 13.82	œ	4038	z	¥	8.669	61.321	40.514		7
z	ILE A	5.070	69.299	33		٦	4039	ర		8.900	60.252	41,495		9
ΰ	ILE A	3.674	70.061	33.192		9	4040	ပ	THR A 516	9.271	60.860	42.835	1.00 11.71	9
ບ	ILE A	2.746	68.964	33	7	9	4041	0	4	10.092		42.959		æ
	ILE A	3.156	67.806	33	13	æ	4042	g	æ	10.107	59.384	41.001	1.00 14.88	9
	ILE A	3.456	70.272	31.683	12	9	4043	ogı	4	9.696		39.742	14	œ
ິບ	ILE A	3.840	69.011	30.868	1.00 11.82	9	4044	8	ø	10.446	58.275	42.004	14	ø
	ILE A	4.241	71.485	31.163	14	9	4045	z	ILE A 517	8.600	60.281	43.863	1.00 9.66	۲
	ILE A	3.293	69.108	29.417	1.00 18.53	ø	4046	ð	ø	8.878		45.251	1.00 10.17	9
z	PRO A	1.462	69.267	33.824	1.00 13.51	7	4047	ပ	ø	9.670	59.586	45.920		9
Ö	PRO A	0.442	68.291	34.164	1.00 14.15	9	4048	0	ILE A 517	9.172	58.427	45.949	13.	80
υ	PRO A	0.531	67.082	33.263	1.00 13.29	9	4049	g	ILE A 517	7.493	60.833	45.948	1.00 12.16	9
0	PRO A	0.780	67.180	32.026	1.00 13.86	œ	4050	g	ILE A 517	6.659	61.984	45.278	1.00 15.38	y
Ü	PRO A	-0.913	69.037	33.810	1.00 13.10	9	4051	CG5	ILE A 517	7.745	61.419	47.392	1.00 14.69	9
ರ	PRO A	-0.528	70.435	34.265	1.00 15.94	9	4052	9	ILE A 517	5.200	61.951	45.779	1.00 22.16	ø
Ħ	CD PRO A 511	0.925	70.625	. 72	1.00 16.03	9	4053	z	ASP A 518	10.911	59.861	46.363	1.00 13.29	7

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40.489	40.679	40.166	38.998	38.950	53	41.415	41.746	40.613	40.252	43.080	43.337	44.226	40.055	39.108	39.732	39.375	38.209	37.246	36.368	34.949	34.672	40.787	41.587	42.999	43.306	40.903	41.043	43.953	45.378	46.053	46.008	46.101	47.502	48.584	48.006	49.715	49.381	47.440	50.201	48.223	49.619
60.043	59.242	59.697	59.955	60.651	58.508	58.176	57.394	56.415	55.599	26.665	55.621	57.646	56.340	55.341	54.044	52.885	55.914	54.859	55.636	55.199	53.976	54.212	53.073	53.502	54.680	52.424	53.317	52.536	52.905	51.785	50.647	53.084	53.605	52.924	54.854	53.673	54.894	55.984	55.983	57.054	57.063
-3.151	-1.865	-0.823	-3.564	-4.828	-3.697	-1.925	-0.704	-0.516	-1.390	-0.896	0.219	-1.016	0.696	1.119	1.626	1.313	2.264	2.814	3.860	3.601	4.369	2.424	2.919	3.231	3.482	4.136	5.270	3.206	3.361	4.148	3.682	2.034	2.124	2.645	1.689	2.542	1.976	1.086	1.703	0.787	1.076
Æ.	THR A	-	THR A	THR A	CG2 THR A 537	N VAL A 538	VAL A	æ	VAL A	CB VAL A 538	VAL A	2 VAL A	ø	LYS A	LYS A	O LYS A 539	LYS A	LYS A	LYS A	LYS A	LYS A		SER A	¥	SER A		SER A	ø	TRP	TRP A	TRP A		TRP A		CD2 TRP A 541	NEI TRP A 541	TRP A	TRP A	-	-	CH2 TRP A 541
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48.090	49.308	48.340	45.569	45.142	45.496	44.989	43.604	43.040	43.191	42.240	42.629	41.691	41.836	46.490	46.884	47.378	47.068	47.921	48.400	47.370	47.637	46.097	44.999	44.894	44.712	43.645	42.509	43.768	44.967	44.890	43.536	42.462	45.192	46.576	45.026	43.562	42.337	42.042	43.030	42.384	40.791
66.621	66.722	65.858	67.691	68.822	70.073	70.292	68.823	67.777	66.431	68.228	65.515	67.306	65.943	70.806	72.077	71.702	72.482	70.510	70.089	69.372	68.817	69.391	68.721	67.275	66.888	69.480	68.739	70.937	66.334	64.927	64.521	64.893	64.042	64.284	62.565	63.773	63.324	61.859	61.111	63.441	61.461
-1.954	-1.209	-3.279	-0.395	0.458	-0.344	-1.454	0.659	1.611	1.438	2.662	2.288	3.545	3.385	0.118	-0.569	-1.992	-2.928	-2.193	-3.524	-4.368	-5.463	-3.923	-4.592	-4.197	-3.019	-4.368	-5.101	-4.807	-5.185	-4.827	-4.271	-4.796	-6.065	-6.446	-5.787	-3.162	-2.521	-2.808	-2.929	-0.976	-2.937
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45.741	45.379	44.044	45.010	43.320	46.821	47.739	47.564	47.579	49.195	49.580	49.201	50.251	47.465	47.392	48.826	49.616	46.555	46.488	45.122	49.097	50.443	50.269	49.522	51.206	52.656	53.523	53.312	54.033	51.047	51.051		53.398	51.036	51.185	49.672	52.140	53.380	53.412	52.456	53.361	53.406
72.927	72.402	73.378	73.153	72.099	72.559	71.569	70.202	70.015	72.027	73.219	73.267	74.127	69.205	67.801	67.207	67.470	66.953	65.496	67.516	66.439	65.835	64.452	64.157	66.720	66.177	67.110	66.912	68.004	63.489	62.108	•	62.324	61.158	59.677	61.228	61.156	60.806	59.308	58.642	61.496	62.891
4.469	5.520	2.616	1.566	2.886	3.750	4.351	3.718	2.469	4.129	4.998	6.174	4.468	4.576	4.161	4.193	5.085	5.144	4.738	5.186		2.989	2.427	1.502	1.960	1.847	1.025	-0.461	-1.198	2.947	2.525	1.751	2.150	3.735	3.312	4.473	0.603	-0.181	-0.261	-0.679	-1.542	-1.367
C THR A 558	THR A	THR A	THR A	THR A	ASP A	ASP A	ASP A	ASP A		ASP A	ASP A	22 ASP A	VAL A	VAL A	VAL A	4	VAL A	VAL A	VAL A	LYS A	LYS A	LYS A	LYS A	CB LYS A 561	LYS A	LYS A	LYS A	LYS A	VAL A	VAL A	•	VAL A	VAL A	VAL A	Ø	-	THR A	THR A		CB THR A 563	OG1 THR A 563
4344	4345	4346	4347	4348	4349	4350	4351	4352	4353	4354	4355	4356	4357	4358	4359	4360	4361							4368							4375		4377	4378	4379	4380	4381	4382	4383	4384	4385
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40.707	39.973	36.821	35.941	36.748	36.917	35.075	33.925	33.553	33.239	37.402	38.346	37.901	36,881	39.803	39.883	41.579	42.426	38.619	38.289	38.321	39.141	39.471	37.500	37.489	38.600	39.204	36.142	38.857	39.947	40.815	40.670	41.670	42.626	43.737	44.188	43.221	44.069	43.141	44.898	43.858	44.818
166.99	66.849		69.248	70.475	70.744	68.759	902.69	70.583	69.481	71.146	72.208	73.456	73.427	71.835	71.490	70.962	72.450	74.532	75.834	75.788	75.032	76.757	76.591	76.567	77.356	78.305	77.200	76.966	77.675	76.639	75.415	77.130	76.232	75.806		76.999	76.123	75.220	77.046	74.500	73.941
-1.772	-0.427	-3.658	-4.017	-4.401	-5.630	-5.198	-5.522	-4.763	-6.635	-3.487	-3.722	-3.003	-2.319	-3.328	-1.826	-1.364	-1.416	-3.278	-2.711	-1.169	-0.631	-3.075	-0.537	0.947	1.568	1.051	1.375	2.874	3.602	4.312	4.121	5.203	5.876	4.918	4.110	7.091	8.018	8.847	8.965	4.801	3.807
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WAT V		88.789	33.415	31.	80	5981	OWO WAT V 539	20.256	45.911	20.702		8
WAT V		41.938	29.865	1.00 32.38	œ	5982	OWO WAT V 540	37.253	43.882	46.170	1.00 33.73	8
WAT V		51.844	54.727	1.00 32.32	œ	5983	OWO WAT V 541	31.158	88.960	12.651	34	60
WAT V		76.377	49.777	1.00 32.31	œ	5984	OWO WAT V 542	2.885	65.252	30.182	1.00 33.66	8
		47.736	42.135		ထ	5882		24.093	72.008	54.068	33	æ
WAT V		46.357	48.051	1.00 32.64	ထ	2986	OWO WAT V 544	-4.660	58.902	52.534	34	8
WAT V		60.897	7.569	1.00 32.20	œ	5987	OWO WAT V 545	20.523	52.571	71.335	34	œ
WAT V		50.290	36.850	1.00 32.13	œ	5988	OWO WAT V 546	50.389	70.373	43.120	34	80
WAT V		67.493	23.991	1.00 32.68	80	5989	OWO WAT V 547	-1.784	74.917	34.717	34	œ
WAT V		70.176	56.941	1.00 32.03	8	2990	OWO WAT V 548	25.051	100.468	31.517	33	60
WAT V		79.662	8.514	1.00 31.76	89	5991	OWO WAT V 549	21.989	83.148	46.194	1.00 34.71	œ
WAT V	-3.757	80.394	22.899	1.00 32.62	œ	5992	OWO WAT V 550	47.521	62.364	11.718	34	ω
WAT V		85.936	45.262		∞	5993	WAT V	52.236	59.305	62.786		80
WAT V		97.650	41.755	1.00 32.31	80	5994	WAT V	40.232	80.510	16.448		8
WAT V		50.551	69.455		80	5995	WAT V	46.253	56.949	70.930	33	8
WAT V		99.962	23.550	1.00 33.21	œ	9669		47.895	53.053	37,454		8
OWO WAT V 513		65.717	41.690		00	5997	WAT V	13.358	71.030	49.662		1:
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OWO WAT V 515		87.045	17.346	1.00 32.99	œ	5999		43.973	74.382	6.399		œ
OWO WAT V 516		54.074	30.752		œ	0009	WAT V	35.593	60.367	77.022		œ
WAT V		85.163	42.409	1.00 32.78	80	6001	WAT V	6.112	83.592	41.192	34	œ
		50.503	27.538	1.00 32.69	80	6002	WAT V	38.614	41.425	25.492	1.00 34.42	80
WAT V	m	91.176	22.385	1.00 33.06	60	6003	OWO WAT V 561	34.074	88.144	12.960	34	æ
OWO WAT V 520	2	41.296	32.218	1.00 34.59	æ	6004	WAT V	40.114	63.213	5.786	34	80
OWO WAT V 521	C)	60.512	23.358		œ	6005	WAT V	-0.202	64.215	60.019	34	8
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WAT V	7	69.394	5.371	1.00 32.65	80	6007	WAT V	15.212	50.738	32.687		80
OWO WAT V 524	7	102.365	27.670	1.00 32.87	ω	6009	OWO WAT V 566	13.018	70.753	53.378		80
OWO WAT V 525	6	48.822	64.451	1.00 32.62	60	6009	OWO WAT V 567	37.836	57.700	75.968	1.00 33.91	89
-	w	46.627	21.339	1.00 33.04	ω	6010	OWO WAT V 568	18.054	71.354	7.047	1.00 34.28	8
WAT V	6	66.472	60.034	1.00 33.02	œ	6011	OWO WAT V 569	20.435	57.091	12.782	1.00 34.06	8
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OWO WAT V 530	m	81.260	18.821	1.00 33.19	œ	6014	OWO WAT V 572	18.083	105.524	25.739	1.00 35.26	8
WAT V	35.980	91.602	25.073	1.00 32.89	œ	6015	OWO WAT V 573	42.363	51.035	61.868	1.00 34.64	æ
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CLAIMS:

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- 1. A method of producing a variant of a parent maltogenic alpha-amylase, said method comprising
 - a) modeling the parent alpha-amylase on the three-dimensional structure of SEQ ID NO: 1 depicted in the Appendix to produce a three-dimensional structure of the parent alpha-amylase;
 - b) identifying in the three-dimensional structure obtained in step (a) at least one structural part of the parent wherein an alteration in said structural part is predicted to result in said altered property;
- modifying the sequence of a nucleic acid encoding the parent alphaamylase to produce a nucleic acid encoding a deletion, insertion, or substitution of one or more amino acids at a position corresponding to said structural part; and
 - d) expressing the modified nucleic acid in a host cell to produce the variant alpha-amylase,

wherein the variant has alpha-amylase enzymatic activity and has at least one altered property relative to the parent.

- The method of claim 1, wherein the altered property is pH dependent activity, thermostability, substrate cleavage pattern, specific activity of cleavage, transglyco-sylation, ability to reduce retrogradation of starch, ability to reduce staling of bread, substrate specificity, substrate binding or calcium binding.
 - 3. A method of constructing a variant of a parent maltogenic alpha-amylase, which method comprises:
 - a) identifying an amino acid residue which is within 15 Å (in particular 10 Å)
 from an active site residue of the parent amylase in the three-dimensional
 structure of said parent, and which is involved in electrostatic or hydrophobic
 interactions with an active site residue;
 - b) substituting said amino acid residue with another amino acid residue which changes the electrostatic and/or hydrophobic surroundings of an active site residue, and which can be accommodated in the structure;
 - c) optionally repeating steps a) and b) recursively;
 - d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or mor positions other than b),

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- e) preparing the variant resulting from steps a) d);
- f) testing the pH dependent activity of said variant; and
- g) optionally repeating steps a) f) recursively; and
- h) selecting a variant having an altered pH dependent activity as compared to the parent amylase.
- 4. A method of constructing a variant of a parent maltogenic alpha-amylase, which method comprises:
 - a) identifying an internal cavity or crevice in the three-dimensional structure of said parent;
 - b) substituting an amino acid residue in the neighborhood of the cavity or crevice with another amino acid residue which increases the hydrophobic interaction and/or fills out or reduces the size of the cavity or crevice;
 - c) optionally repeating steps a) and b) recursively;
- d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),
 - e) preparing the variant resulting from steps a) d);
 - f) testing the thermostability of said variant; and
 - g) optionally repeating steps a) f) recursively; and
 - h) selecting a variant having increased thermostability as compared to the parent amylase.
- 5. The method of claim 4, wherein the substitution of the amino acid residue results in increasing the hydrophobic interaction, substitution with proline, substitution of histidine with another amino acid, stabilization of calcium binding, introduction of an interdomain disulfide bond, removal of a deamidation site, altering a hydrogen bond contact, filling in an internal structural cavity with an amino acid with a bulkier side group, introduction of interdomain interactions, altering charge distribution, helix capping, or introduction of a salt bridge.
 - 6. A method of constructing a variant of a parent maltogenic alpha-amylase, which method comprises:
 - a) identifying an amino acid residue within 10 Å from a calcium binding site in the three dimensional structure of the amylase;
 - b) substituting the amino acid residue with another amino acid residue so as to improve the interaction with the calcium ion;
 - c) optionally rep ating steps a) and b) recursively;

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- d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),
- e) preparing the variant resulting from steps a) d);
- f) testing the thermostability of said variant; and
- g) optionally repeating steps a) f) recursively; and
 - h) selecting a variant having increased thermostability as compared to the parent amylase.
- 7. A method of constructing a variant of a parent maltogenic alpha-amylase, which method comprises:
- a) identifying the substrate binding area in a model of the three-dimensional structure of the parent amylase;
 - b) modifying the substrate binding area by an amino acid substitution, deletion or insertion;
 - c) optionally repeating step b) recursively;
- d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),
 - e) preparing the variant resulting from steps a) d);
 - f) testing the substrate-cleavage pattern of the variant.
 - g) optionally repeating steps a) f) recursively; and
- 20 h) selecting a variant having an altered substrate-cleavage pattern as compared to the parent amylase.
 - 8. A method for producing a maltogenic alpha-amylase variant, comprising:
 - a) constructing the variant by the method of any of claims 2-7;
 - b) transforming a microorganism with a DNA sequence encoding the variant;
- c) cultivating the transformed microorganism under conditions which are conducive for producing the variant, and
 - d) optionally, recovering the variant from the resulting culture broth.
 - 9. A polypeptide which:

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- a) has maltogenic alpha-amylase activity;
- b) has at least 70 % identity to SEQ ID NO: 1;
 - c) comprises an amino acid modification compared to SEQ ID NO: 1 at a position corresponding to D127, V129, F188, A229, Y258, V281, F284, T288, N327, M330, G370, N371, and/or D372; and
 - d) has altered pH dependent activity as compared to the polypeptide of SEQ
 ID NO: 1.

- 10. The polypeptide of claim 9, wherein the modification comprises a substitution corresponding to D127N/L, V129S/T/G/V, F188E/K/H, A229S/T/G/V, Y258E/D/K/R/F/N, V281L/T, F284K/H/D/E/Y, T288E/K/R, N327D, M330L/F/I/D/E/K, G370N, N371D/E/G/K, and/or D372N/V.
- 5 11. A polypeptide which:
 - a) has maltogenic alpha-amylase activity;
 - b) has at least 70 % identity to SEQ ID NO: 1; and
- c) comprises an amino acid modification compared to SEQ ID NO: 1 at a position corresponding to Q13, I16, D17, N26, N28, P29, A30, S32, Y33, G34, L35, K40, M45, P73, V74, D76, N77, D79, N86, R95, N99, I100, H103, Q119, N120, N131, S141, T142, A148, N152, A163, H169, N171, G172, I174, N176, N187, F188, A192, Q201, N203, H220, N234, G236, Q247, K249, D261, N266, L268, R272, N275, N276, V279, N280, V281, D285, N287, F297, Q299, N305, K316, N320, L321, N327, A341, N342, A348, Q365, N371, N375, M378, G397, A381, F389, N401, A403, K425, N436, S442, N454, N468, N474, S479, A483, A486, V487, S493, T494, S495, A496, S497, A498, Q500, N507, I510, N513, K520, Q526, A555, A564, S573, N575, Q581, S583, F586, K589, N595, G618, N621, Q624, A629, F636, K645, N664 and/or T681; and
 - d) has improved stability compared to the polypeptide of SEQ ID NO: 1.
- The polypeptide of claim 11, wherein the modification comprises at a position corresponding to K40, V74, H103, S141, T142, F188, H220, N234, K249, D261, L268, V279, N342, H344, G397, A403, K425, S442, S479, S493, T494, S495, A496, S497, A498, Q500, K520, A555 and/or N595; preferably a substitution corresponding to K40R, V74P, H103Y/V/I/L/F/Y, S141P, T142A, F188I/L, H220Y/L/M, N234P, K249P, D261G, L268P, V279P, N342P, H344E/Q/N/D/Y, G397P, A403P, K425E, S442P, S479P, S493P, T494P, S495P, A496P, S497P, A498P, Q500P, K520R, A555P and/or N595I.
- The polypeptide of claim 11 or 12, wherein the modification comprises at a position corresponding to D17, N28, P29, A30, S32, Y33, G34, R95, H103, N131, H169, I174 and/or Q201 such as to improve calcium coordination, preferably a substitution corresponding to D17Q/E, A30D/M/L/AV/I/E/Q, S32D/E/N/Q, R95M/L/AV/I/E/Q, H103Y/N/Q/D/E, N131D, H169N/D/E/Q, I174E/Q, Q201E.
 - 14. The polypeptide of any of claims 11-13, wherein the modification comprises a substitution at a position corresponding to Q13, N26, N77, N86, N99, Q119, N120,

N131, N152, N171, N176, N187, Q201, N203, N234, Q247, N266, N275, N276, N280, N287, Q299, N320, N327, N342, Q365, N371, N375, N401, N436, N454, N468, N474, Q500, N507, N513, Q526, N575, Q581, N621, Q624 and/or N664 such as to remove a deamidation site, preferably a substitution corresponding to 5 Q13S/T/AV/L/I/F/M, N26S/T/AV/L/I, N77S/T/AV/L/I, N86S/T/AV/L/I, N99T/S/V/L, N120S/T/A/V/L/I, N131S/T/A/V/L/I, N152T/S/V/L, N171Y/D/S/T. Q119T/S, N187S/T/A/V/L/I, Q201S/T/A/V/L/I/F/M, N203D/S/T/A/V/L/I. N176S/T/A/V/L/I. Q247S/T/A/V/L/I/F/M, N266S/T/A/V/L/I, N234S/T/A/V/L/I. N275S/T/A/V/L/I, N276S/T/A/V/L/I, N280S/T/A/V/L/I, N287S/T/A/V/L/I, Q299L/T/S, N320S/T/A/V/L/I, N342S/T/A/V/L/I, Q365S/T/A/V/L/I, N371S/T/A/V/L/I. 10 N327S/T/A/V/L/I, N401S/T/A/V/L/I, N436S/T/A/V/L/I, N454D/S/T/A/V/L/I, N375S/T/A/V/L/I, N474D/S/T/A/V/L/I, Q500S/T/A/V/L/I/F/M, N507S/T/A/V/L/I, N468D/S/T/A/V/L/I. N513S/T/A/V/L/I, Q526 D/S/T/A/V/L/I, N575S/T/A/V/L/I, Q581S/T/A/V/L/I/F/M, N621S/T/AV/L/I Q624S/T/AV/L/I/F/M and/or N664D/S/T/AV/L/I.

- The polypeptide of any of claims 11-14, wherein the modification comprises a substitution at a position corresponding to I16, L35, M45, P73, D76, D79, A192, I100, A148, A163+G172, L268, V281, D285, L321, F297, N305, K316, S573, A341, M378, A381, F389, A483, A486, I510, A564, F586, K589, F636, K645, A629, and/or T681 such as to improve hydrogen bond contact, preferably a substitution corresponding to I16T/D/N, L35Q, M45K, P73Q, D76E, D79E/Y, A192S/D/N, I100T/S/D/N/E/Q, A148D/N/E/Q/S/T/R/K, A163Y+G172S/D/N, L268R/K, V281/Q, D285R/K, L321Q, F297N/D/Q/E, N305K/R, K316N/D, S573N/D, A341R/K, M378R/K, A381S/D/N, F389Y, A483S/D/N, A486Q/E, I510R/K, A564S/D/N, F586S/D/N, 'K589S/D/Q/N, F636Y, K645T, A629N/D/E/Q, and/or T681D/N/E/Q/S.
- 25 16. The polypeptide of any of claims 11-15, wherein the modification comprises substitutions such as to introduce one or more interdomain disulfide bonds, preferably corresponding to G236C + S583C, G618C + R272C, and/or A348C + V487C.
- The polypeptide of any of claims 11-16, wherein the substitution at a position corresponding to L51, L75, L78, G88, G91, T94, V114, I125, V126, T134, G157, L217, S235, G236, V254, V279, V281, L286, V289, I290, V308, L321, I325, D326, L343, F349, S353, I359, I405, L448, Q449, L452, I470, G509, V515, S583, G625, L627, L628 and/or A670 so as to fill an internal cavity or crevice, preferably a substitution corresponding to L51W, L75F/Y, L78I, G88A/V/T, G91T/S/V/N, T94V/I/L, V114V/I/L, I125L/M/F/Y/W, V126I/L, T134V/I/L/M/F/Y/W, G157A/V/I/L,
 L217V/I/M/F/Y/W, S235I/L/M/F/Y/W, G236A/V/I/L/M/F/Y/W, V254I/L/M/F/Y/W,

V279M/I/L/F, V281I/L/M/F/Y/W, L286F, V289I/L/R, I290M/L/F, V308I/L/M/F/Y/W, L321I/M/F/Y/W, I325L/M/F/Y/W, D326E/Q, L343M/F/Y/W, F349W/Y, S353V/I/L, I359L/M/F/Y/W, I405M/L/Y/F/W, L448Y, Q449Y, L452M/Y/F/W, I470M/L/F, G509A/V/I/L/M/S/T/D/N, V515I/L, S583V/I/L/V, G625A/V/I/L/M/F/Y/W, L627M/F/Y, L628M/I/F/Y/W, A670V/I/L/M/F/Y/W and/or L217 in combination with L75 (e.g. L217F/Y in combination with L75F/Y).

- 18. The polypeptide of any of claims 11-17, wherein the modification comprises a substitution at a position corresponding to N106, N320 and Q624 so as to create a salt bridge, preferably a substitution corresponding to N106R, N320E/D and/or 10 Q624E.
 - 19. The polypeptide of any of claims 11-18, wherein the modification comprises a substitution at a position corresponding to K244 and/or K316 such as to alter the charge distribution, preferably a substitution corresponding to K244S and/or K316G/N/D.
- 15 20. The polypeptide of any of claims 11-19, wherein the modification comprises a substitution at a position corresponding to V281 and/or A629 such as to alter the binding site, preferably a substitution corresponding to V281Q and/or A629N/D/E/Q.
- 21. The polypeptide of any of claims 11-20, wherein the modification comprises substitutions such as to alter the interdomain interaction at a position corresponding to F143+F194+L78, A341+A348+L398+l415+T439+L464+L465, L557, S240+L268, Q208+L628, F427+Q500+N507+M508+S573 and/or I510+V620, preferably substitutions corresponding to F143Y+F194Y+L78Y/F/W/E/Q, A341S/D/N+A348V/I/L+L398E/Q/N/D+I415E/Q+T439D/E/Q/N+L464D/E+L465D/E/N/Q/R/K, L557Q/E/N/D, S240D/E/N/Q+L268D/E/N/Q/R/K, Q208D/E/Q+L628E/Q/N/D, F427E/Q/R/K/Y+Q500Y+N507Q/E/D+M508K/R/E/Q+S573D/E/N/Q; and/or I510D/E/N/Q/S+V620D/E/N/Q.
 - 22. A polypeptide which:
 - a) has maltogenic alpha-amylase activity;
 - b) has at least 70 % identity to SEQ ID NO: 1;
- c) comprises an amino acid modification compared to SEQ ID NO: 1 at a position corresponding to P191, A192, G193, F194 and/or S195; and
 - d) has higher specific amylase activity than the polypeptide of SEQ ID NO:
 - 1.

- 23. The polypeptide of claim 22, wherein the modification comprises a deletion, preferably the deletion Δ (191-195).
- 24. The polypeptide of claim 22, wherein the modification comprises insertion, preferably 192-A-193.
- 5 25. A polypeptide which:

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- a) has maltogenic alpha-amylase activity;
- b) has at least 70 % identity to SEQ ID NO: 1;
- c) comprises an amino acid modification compared to SEQ ID NO: 1 at a position corresponding to A30, K40, N115, T142, F188, T189, P191, A192, G193, F194, S195, D261, T288, N327, K425, K520 and/or N595; and
- d) has a higher ability than the polypeptide of SEQ ID NO: 1 to reduce retrogradation of starch and/or staling of bread.
- 26. The polypeptide of claim 26, wherein the modification comprises A30D, K40R, N115D, T142A F188L, T189Y, Δ (191-195), D261G, T288P, N327S, K425E, K520R and/or N595I.
 - 27. A nucleic acid sequence encoding the polypeptide of any of claims 9-26, preferably operably linked to one or more control sequences which direct the expression of the variant in a suitable expression host.
- 28. A recombinant expression vector comprising the nucleic acid sequence of claim 27, a promoter, and transcriptional and translational stop signals, and preferably further comprising a selectable marker.
 - 29. A transformed host cell comprising the nucleic acid sequence of claim 27 or the vector of claim 28.
 - 30. A method for producing the polypeptide of any of claims 9-26, comprising:
- a) cultivating the transformed host cell of claim 29 under conditions conducive to expression of the variant; and
 - b) recovering the variant.
 - 31. A process for preparing a dough or a baked product prepared from the dough which comprises adding the polypeptide of any of claims 9-26, or a variant produced

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by the method of any of claims 1-8 to the dough in an amount which is effective to retard the staling of the bread.

32. The process of claim 31, wherein the variant is added in an amount of 0.1-5 mg per kg of flour, preferably 0.5-2 mg/kg.

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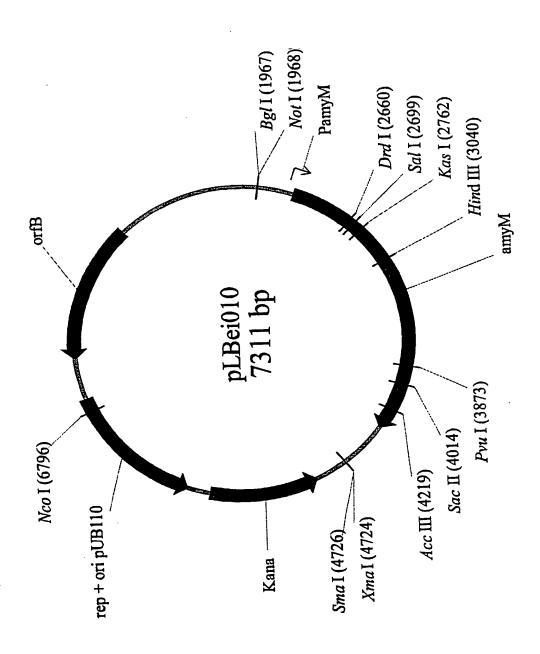


Fig. 1

PCT/DK99/00088

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Abb	
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~-+			~a~	2++	asa	maa	cat	ttc	aaa	aat	taa	acc	aca	+++	gac	432
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АВР	Pne	пув	GIII	100	GIU	GIU	HID	FILE	105	YOU	11.0			110	A D D	
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Thr	Leu	vaı		Asp	АТА	HIS	GIN		GIĀ	Ile	гув	VAI		VAI	Авр	
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y	225	_	~-0			230	_		- 110	*****	235	_			-,,,	

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INTERNATIONAL SEARCH REPORT

International application No. PCT/DK 99/00088

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A. CLASSIFICATION OF SUBJECT MATTER		
IPC6: C12N 9/28, A21D 8/04		
According to International Patent Classification (IPC) or to both no B. FIELDS SEARCHED	ational classification and IPC	
Minimum documentation searched (classification system followed by	v classification symbols)	
IPC6: C12N	,,	
Documentation searched other than minimum documentation to the	extent that such documents are include	ed in the fields searched
SE,DK,FI,NO classes as above		
Electronic data base consulted during the international search (name	of data base and, where practicable, se	arch terms used)
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
		1
Category* Citation of document, with indication, where app	propriate, of the relevant passages	Relevant to claim No.
X WO 9623874 A1 (NOVO NORDISK A/S) (08.08.96), see page 30-37,	, 8 August 1996 claims 1-2	1-32
X WO 9743424 A1 (GENENCOR INTERNAT 20 November 1997 (20.11.97) 21-34, page 12, lines 15-19		1-8
		7 22
A		7-32
X WO 9741213 A1 (NOVO NORDISK A/S) (06.11.97), see page 11, lii), 6 November 1997 nes 5-12	1-8
A		9-32
X Further documents are listed in the continuation of Box	C. X See patent family an	nex.
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance.	"T" later document published after the date and not in conflict with the a the principle or theory underlying	
"E" ertier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other	"X" document of particular relevance: considered novel or cannot be cor step when the document is taken a	adered to involve an inventive
special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means	"Y" document of particular relevance: considered to involve an inventive combined with one or more other	such documents, such combination
"P" document published prior to the international filing date but later than the priority date claimed	being obvious to a person skilled a "&" document member of the same pa	
Date of the actual completion of the international search	Date of mailing of the internation	
8 June 1999	0 8 -06- 1999	
Name and mailing address of the ISA/	Authorized officer	
Swedish Patent Office Box 5055, S-102 42 STOCKHOLM	Yvonne Siösteen	
Facsimile No. + 46 8 666 02 86	Telephone No. + 46 8 782 25 6	00

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Form PCT/ISA/210 (second sheet) (July 1992)

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International application No.

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WO	9623874	A1	08/08/96	AU BR CA CN EP JP	4483496 9607013 2211316 1172501 0808363 11500003	A A A	21/08/96 28/10/97 08/08/96 04/02/98 26/11/97 06/01/99
0	9743424	A1	20/11/97	AU US	2996997 5763385		05/12/97 09/06/98
10	9741213	A1	06/11/97	AU EP	2692897 0904360		19/11/97 31/03/99